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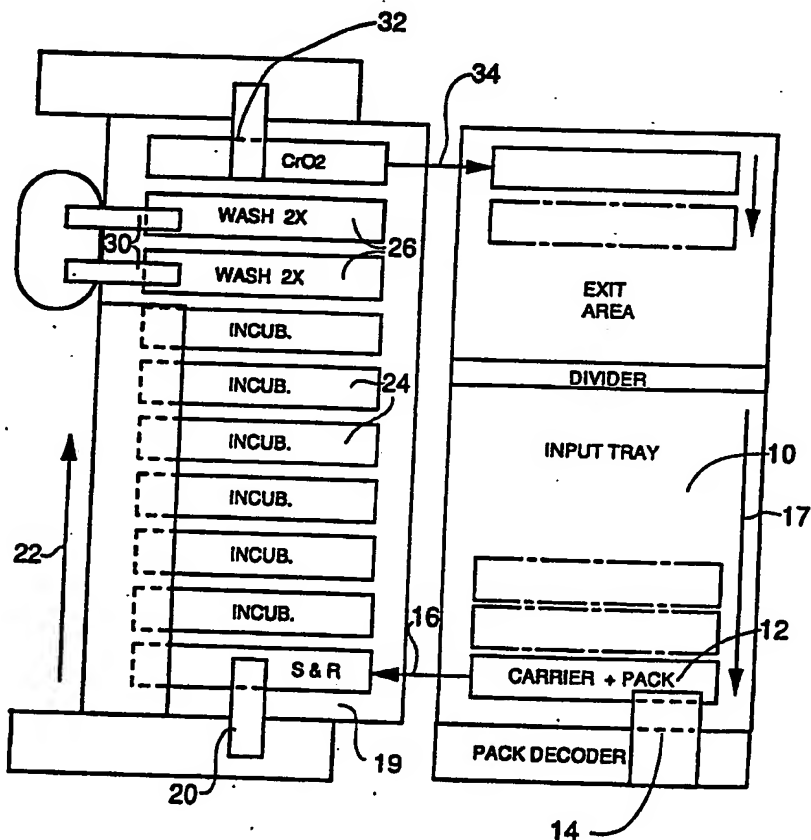
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(54) Title: MULTI-LINEAR AUTOMATIC APPARATUS FOR PROCESSING IMMUNOASSAYS

(57) Abstract

The apparatus processes immunoassays of samples using a solid support. It provides a compact system using rectilinear motion to process carriers holding samples on a real time basis while permitting the input and output of the system to operate on a random basis.



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Title

Multi-Linear Automatic Apparatus
for Processing Immunoassays

5 Cross Reference to Related Applications

Subject matter disclosed herein is disclosed or
claimed in copending applications Method and Apparatus
for Automatically Processing Magnetic Solid Phase
Reagents, filed August 10, 1988, Serial No. 07/230,449
10 (IP-0754), Method and Apparatus for Effecting the
Automatic Analytical Testing of Samples, filed
August 26, 1988, Serial No. 07/737,119 (IP-0751), Vortex
Mixer Drive, filed 26 July 1991, Serial No. 07/736,177
(IP-0801), and Carrier Device, filed 26 July 1991, Serial
15 No. 07/736,155 (IP-0911).

Field of the Invention

The present invention relates to an automated
apparatus for analyzing liquid samples, and, more
20 particularly, to apparatus for processing samples for
quantitatively determining the presence of a variety of
substances, including antibodies, antigens and various
proteins, including cancer markers and hormones, in
biological samples.

25

Background of the Invention

Heterogeneous immunoassays typically are performed
using a solid support, preferably with the solid support
being formed from magnetic particles. A particularly
30 preferred support for this purpose is that described in
U.S. Patent 4,661,408 assigned to E. I. du Pont
de Nemours & Co., Inc. This patent describes a chromium
dioxide particle which has favorable magnetic properties
for use as a solid support in such assays.

The concept of using magnetically responsive particles to effect separations of bioactive materials is old in the art (Hedin, C. G., Biotech. Bioeng. Symp. No. 3 (1972) 173-174; Robinson, P. J., et al., Biotech Bioeng. (1973) 15, 603-606). The concept has been extended over time to include affinity purification of enzymes, proteins or microorganisms applicable to any sorption-desorption process (Dunhill, P., et al., Biotech. Bioeng. (1974) 10, 987-990; Horisberger, M., Biotech. Bioeng. (1976) 18, 1647-1651).

Another improved magnetically responsive particle is described by Mansfield et al. in U.S. Patent No. 4,197,337. These particles are porous glass microparticles with magnetic material imbedded within them. This gives the particles the properties of high surface area, inertness and being substantially superparamagnetic. This high surface area again favors rapid reaction kinetics and increases capacity of the individual particles. Being substantially superparamagnetic, basically means the particles do not retain much magnetic memory, or retentivity, when removed from a magnetic field. This means that particles can be repeatedly separated from their environment by a magnetic field without affecting the ability to redisperse those particles. This is of advantage in sandwich immunoassays where multiple washing steps may require repeated separation and redispersion.

The protected CrO_2 particles described in the Du Pont patent have several properties that are particularly advantageous in heterogeneous immunoassays. These are:

low remanent magnetism and favorable surface structure -- allowing repeated magnetic separation/dispersion cycles;
rapid separation in a magnetic field;
5 high surface area for high capture capacity;
a highly stable particle for maximum reagent shelf life.

One problem with immunoassays is that they require repeated washing of the solid support containing the
10 bound component following removal of the free component. This is a particularly difficult procedure even when performed manually. It is a particular problem when the automatic wash procedure is incorporated into an automatic instrument which is capable of performing
15 immunoassays. The typical requirements for purity in such immunoassays is that the bound component remaining following such washes should not contain more than 20 parts per million of the original sample/conjugate matrix. This necessitates the use of multiple wash
20 stations and can be accomplished in automatic instruments by providing such multiple wash stations. However, the several mechanisms required to operate individually the several wash stations and the mechanisms required for the wash stations themselves can
25 become quite expensive.

There are essentially two types of heterogeneous immunoassays. These are competitive immunoassays and sandwich immunoassays. In a competitive assay, an
30 antibody to an antigen contained in a first reagent is attached to the derivatized magnetic particles to make up a solid phase. The second reagent, consisting of antigen attached to a tag (a measurable entity, including radioactive molecules, fluorescent molecules, or enzymes), and patient sample are mixed with the solid

phase in a test tube. In the absence of patient antigen, some 50% of the antigen-tag is bound to the antibody of the magnetic solid phase. In the presence of patient antigen, some of the antibodies are filled up
5 with patient antigen and are unavailable to the tag antigen. As a result, increasing amount of patient antigen leads to decreasing amount of tag antigen. Thus one can form a calibration chart relating the amount of patient antigen to the amount of tag antigen. The
10 separation and wash stages results from the need to measure the free tag or the bound tag, not the total tag added. The magnetic particle facilitates this separation by forming the particles with the bound tag into a pellet on the side of the tube. The free tag can
15 then be removed as by aspiration. Following the separation and removal of free tag, another reagent is added so that the amount of bound tag can be measured. In a typical case, enzyme is used as the tag so that the reagent added is a substrate for the enzyme permitting
20 the measurement of the amount of tag that was bound to antibody.

In a typical sandwich immunoassay, an antibody to an antigen is attached to the magnetic particle. This is in high concentration relative to the amount of
25 patient antigen in a sample. Patient antigen is captured by the antibody on the magnetic particles and then the particles (and captured patient antigen) separated from interfering substances in the sample. To this, a second reagent, containing a second antibody
30 with an attached tag, is added. This second antibody attaches to the patient antigen, captured by the first antibody on the magnetic particle, and results in the formation of a sandwich so that the second antibody tag is held firmly by the antigen to the first antibody on

the magnetic particle. At this point, a magnetic separation similar to that described, permits the determination of bound tag which is in proportion to the patient antigen, the excess tag of the second reagent having been removed by aspiration.

Magnetic particles are particularly useful as the solid support in heterogeneous immunoassay because they can readily separate the free from the bound tag. Such immunoassays using magnetic particles as a solid support are described for example in U.S. Patent 4,661,408 (Lau et al.), U.S. Patent 4,628,037 issued to Chagnon et al., U.S. Patent 4,672,040 issued to Josephson, and U.S. Patent 4,698,302 issued to Whitehead et al. The methods disclosed in all of these patents separation units such as those that are available from Corning Medical, Corning Glass Works, Medfield, Mass. Such manual techniques are relatively slow, require relatively strong magnets which are expensive, require considerable manual dexterity, and require an excessive amount of time to effect the separation with the purity required, particularly for sandwich type heterogeneous immunoassays.

There are numerous automatic clinical analysis type instruments available on the market today. Typical of these is the Automatic Clinical Analyzer known as the aca® sold by E. I. du Pont de Nemours & Co., Inc. of Wilmington, Delaware. This is an instrument in which an incubator is used to process samples. The incubator is in the form of a belt or chain in which a sample mixed with reagents in packs and is analyzed for its various components. While quite satisfactory for many purposes it does not have the versatility required for some of the more recently developed sensitive immunoassays

Other analyzers that are available for analyzing samples are those that are described, for example, in U.S. Patent 4,315,891 assigned to Olympus Optical Company. This patent describes a belt or chain
5 incubator and has a single reaction line in which reaction vessels are carried step by step along the reaction line. Samples and reagents are delivered to the vessels during their movement along the reaction line to obtain a test liquid which is then subjected to
10 photometric analysis. This machine permits several tests to be run simultaneously on samples by an interleaving processing. Unfortunately, this apparatus does not have the capability for performing the precise washing required in heterogeneous immunoassays. The
15 apparatus also requires a relatively large amount of space.

Another automatic analytical apparatus is that described in U.S. Patent 4,459,265 assigned to Clinicon. This apparatus includes a stepwise rotatable circular
20 plate. It carries a plurality of reaction tubes on its periphery with several reagent supply stations arranged at different locations around such periphery. The use of multiple stations does provide the machine with the versatility to carry out several different test
25 methodologies, but again does not provide the necessary washing required for the more sensitive heterogeneous immunoassays.

Many of the machines described use circular trays that rotate to provide the necessary random access to
30 add sample and reagent to, incubate, wash the magnetic particles, etc., in the reaction vessels. This creates the need for greater space requiring larger instruments that would otherwise be desirable. It is also necessary in such instruments to have random access of the samples

to be processed to the real time of the immunoassay system.

Summary of the Invention

5 Many of these problems of the prior art apparatus for performing immunoassays, including those which use magnetic particles, i.e., particles that are responsive to a magnetic field, are reduced using the apparatus of this invention. The apparatus permits immunoassays to
10 be processed, using such magnetic particles, and yet is compact.

This invention provides an automatic multilinear apparatus for processing immunoassays of samples using a solid support, the assays having bound and free phases,
15 the bound phase being bound to the solid support, the apparatus comprising: an inlet chamber, a plurality of carriers positioned in the inlet chamber, each carrier holding a sample, a rotatably mounted reaction vessel, particles responsive to a magnetic field, reagents, and
20 a reaction product container, a processing chamber generally parallel to the inlet chamber, first means to transport the carriers linearly in a first direction to one end of the inlet chamber, second means to transport sequentially the carriers linearly in a second direction
25 transverse to the first direction from the one end of the inlet chamber to one end of the processing chamber, first translating means for acting on each carrier at the one end of the processing chamber to transfer each carrier's sample and reagents into the carrier's
30 reaction vessel, third means to transport sequentially the carriers linearly in a third direction opposite to and generally parallel to the first direction to several processing positions, means for vortexing at at least one processing position by nutating the lower portion of

each carrier reaction vessel, wash means at at least one processing position for removing liquids from each carrier's reaction vessel and replacing the liquids with a different liquid, magnet means at each wash means
5 position for applying a magnetic field to each carrier reaction vessel prior to liquid removal, second translating means at the last sequential processing position for transferring the contents of each carrier's reaction vessel to its container, fourth means to
10 transport each carrier transversely of the third direction from the processing chamber to the other end of the inlet chamber, and fifth means to transport each carrier from the other end of the inlet chamber in a direction generally parallel to the first direction for
15 storage.

In a preferred embodiment of the invention, each carrier is provided with at least a pair of receptacles positioned at the bottom of the carrier, and the second means has a pair of spring loaded pins foldable
20 downwardly in the second direction, whereby the second means can be displaced in a direction opposite the second direction to engage each pair of receptacles and transport each carrier in the second direction.

In another embodiment of the invention, the third
25 means includes pairs of pins fixed adapted to engage each of the plural carrier receptacles and rectilinear transport means for raising each of the pair of fixed pins to engage the receptacles and displace the carrier's one processing position, and lowering each of
30 the pairs of fixed pins to disengage the receptacles.

In still another embodiment of this invention the magnet means includes a magnet, and means to displace the magnet to engage the side of a reaction vessel, thereby to apply the magnetic field to the particles.

This apparatus permits the apparatus to be used for heterogeneous immunoassays and require only a limited amount of space and yet permits random access to a real time immunoassay process.

5

Brief Description of the Drawings

The invention will be better understood with reference to the several drawings in which like reference numerals are used to indicate like components, in which:

10

Figure 1 is block diagram of an automatic multilinear apparatus for processing immunoassays of samples using magnetic particles as solid support in accordance with this invention;

15

Figure 2 is a pictorial view of the automatic multilinear apparatus of Figure 1 constructed in accordance with this invention;

Figure 3 is a plan view of the apparatus depicted in Figure 2, partially broken away, to reveal the inner portion of the apparatus;

20

Figure 4 is a section line taken along the 4-4 of Figure 3 to illustrate a side elevational view of the apparatus of this invention;

Figure 5 is a sectional view taken along the section line 5-5 of Figure 3 to depict the rectilinear, position by position advance mechanism for the carriers of this invention;

25

Figure 6 is a view taken along the section line 6-6 of Figure 3 to depict the mixer that is used to vortex the reaction vessels of this invention;

30

Figure 7 is a view taken along the section line 7-7 of Figure 3 to depict a wash station of this invention;

Figure 8 is a view taken along the section line 8-8 of Figure 3 to depict the mechanism which propels the carrier;

Figure 9 is exploded view showing the features of
5 the carriers used in this invention;

Figure 10 is a sectional view taken along the section line 10-10 of Figure 9;

Figure 11 is a sectional view taken along the section line 11-11 of Figure 9;

10 Figure 12 is a liquid flow diagram depicting the liquid flows taking place during sample processing in the reaction vessel; and

Figures 13 through 13 are flow charts of the software used to operate the apparatus of this
15 invention.

Detailed Description of the Preferred Embodiment

As may be seen in Figure 1, there is a block diagram illustrating an analyzer in which the apparatus
20 of this invention. The analyzer has an input or inlet chamber 10 in which there are held a plurality of carriers 12. Each carrier 12 holds a sample holder for a sample to be processed, a flexibly mounted reaction vessel. There are magnetic particles in the reaction
25 vessel, i.e., particles that are responsive to a magnetic field, along with reagents for performing immunoassays. The carrier also holds a transparent removable container that is introducible into another instrument for evaluating the processed immunoassay
30 reaction. An optical decoder 14 is positioned at the lower portion of the inlet chamber (in the drawing) for decoding a code on the sample holder in the carrier which will identify the particular test to be run so that the analyzer may be actuated properly.

A first transport 18 moves the carrier 12 linearly in a first direction and urges the carriers against the decoder 14. Once decoded, a shuttle mechanism 16 transports the carriers linearly in a second direction transverse to the first direction from one end of the inlet chamber to one end of a processing chamber 19. The processing chamber 19 is generally parallel to the inlet chamber and is so positioned to minimize the space occupied by the entire processing unit. A first translator symbolized by the block 20 acts on each carrier 12 to transfer each carrier's sample and reagents into the carrier's reaction vessel. A transport mechanism 22 sequentially transports the carriers 12 linearly in a third direction opposite to and generally parallel to the first direction to several processing positions 24. At the second, fourth and sixth processing positions 24 are means of vortexing each carrier's reaction vessel. At the eighth and ninth processing positions, wash means 26 are provided for removing liquids from each carrier reaction vessel and replacing the liquids with a different liquid. Translating means 30 are provided at each wash position to apply a magnetic field to each carrier reaction vessel prior to removal of the liquid so as to position the magnetic particles against walls of the reaction vessel.

Finally, a second translating means 32 at the last processing position transfers the contents of each carrier's reaction vessel into its container for storage until analyzed. A fourth transport means 34 transports each carrier 12 transversely of the third direction from the processing chamber back into the other end of the inlet chamber. Finally, a fifth means 36 operates to transport each carrier 12 from the other end of the

inlet chamber 10 in a direction generally parallel to the first direction for storage. Following storage, the carriers 12 may be removed at will so that the container can be removed therefrom and put into a suitable
5 analyzer for analysis such as the aca® Clinical Analyzer sold by E. I. du Pont de Nemours and Company, Wilmington, DE.

The multilinear apparatus provided thus is seen to provide a significant space saving and yet permits the
10 apparatus to operate randomly against the real time requirements of a processing system for analyzing samples, i.e., it permits randomly entering samples into and removing samples from the system as desired.

There may be seen in Figure 2 a pictorial view of
15 the instrument depicted functionally in Figure 1. Thus in Figure 2 there is an inlet chamber 10, carrier 12, first transport mechanism 16, detector 14, processing chamber 19, and rectangular motion transport mechanism 22. The unit also includes, typically in the box
20 portion 500 (Fig. 3), the electronics and software that control the operation of the system as will be described hereinafter.

In Figures 3 to 12 more detail of the system depicted in Figures 1 and 2 is shown. Thus in these
25 figures there may be seen the first transport means or carrier drive 17 connected to be driven by a spring loaded belt 152 which is actuated by a motor 54. The carrier drive 17 is guided in its operation by guide bars 156. The first transport mechanism 16 (Figure 4)
30 comprises a pair of pins 60 which are spring loaded so as to be foldable in a first direction, i.e., the direction necessary to move the carriers 12 from the inlet chamber 10 to the processing chamber 19. The pins 60 themselves are mounted on a bracket 62 which is

mounted to be supported by bars 65 and a drive belt 66 which provides force for movement. The drive belt 66 is driven by a motor 68.

In their operation, the pins 60 of the first transport mechanism 16 are allowed to move to the right in the drawing, against the first direction, until the pins 60 engage receptacles 70 formed in the bottom of each carrier. Thus engaged, when the movement of the transport mechanism is reversed to move in the first direction, the engaged carrier 12 is moved to the left in the drawing in the first direction into the processing chamber.

The processing chamber 19 includes a platform 67 having a plurality of pins 18 positioned thereon located at the spacing of each of several processing positions. In this illustration, the processing positions total 10 during which the reagents and sample in the reaction vessel will incubate, be washed and then moved out for further processing. The platform 67 is moved in a rectilinear motion so as to move forward in the direction depicted by the arrows 70 from one processing position to the next at intervals. Following each position movement, the platform 66 is lowered as indicated by the arrow 73 and dotted pins 18" (Figure 5) so as to disengage from the several carriers. It is then moved backward against the arrow 70 (Figure 3) to the next preceding processing position and then again raised as shown by arrow 69 and dotted pins 18" (Figure 5) to engage the pins in the carriers at each preceding processing position. In this manner the carriers are moved incrementally from one processing position to the next, to the next and so on. As the carriers 12 move to each processing position since the reaction vessel 100 in each case contains magnetic particles which tend to

settle they must be occasionally vortexed to resuspend the particles. This is accomplished by the mixer at processing position 2, 4, 6, 8, 9, and 10 depicted in Figure 6. The carrier 12 for holding the reaction vessel is best seen in Figs. 9-11.

There may be seen in Figures 9, 10 and 11 an exploded sectional view of one of the carriers 12 of Figure 1. The carrier is seen to contain a hollow, molded housing 250 defined by a pair of sidewalls 52, a top plate 258, and a base support 260. A drive bar 240 is positioned in the lower portion between the sidewalls 252 and is secured to the base support as by gluing. This bar has receptacles to facilitate its receiving drive pins for positioning the bar 240 and hence the carrier. The housing may be formed of polysulfone or any other suitable engineering plastic which is rigid, strong and chemically inert. Attached to the front sidewall (in the drawing) is a partition 254 which cooperates with the top 258 to accommodate the top frame of an analytical pack 262 of an analytical pack 264 which may be the same and preferably is the same as the aca® pack used in the aca® Automatic Clinical Analyzer sold by E. I. du Pont de Nemours and Company, Wilmington, Delaware U.S.A. The aca® pack has identifying indicia 266 on the top which may be read by appropriate sensors to indicate the particular test being run and includes a septum 68 with an orifice 270 which may be used to introduce materials into a plastic pack 72. Since the aca® pack is well known it will not be described further.

In any event, the partition 254 and top 258 cooperate to define an orifice 256 adapted to accommodate the top member of the aca® pack 262 so it may be inserted into the carrier with the lower side

pack 272, which is formed of plastic material. The side pack is to slide in between the two walls 252. The top of the carrier 250 also includes an elongated cup-like member 276 which is adapted to receive a removable sample reservoir 278 containing a reservoir 280. The sample reservoir 278 is held in the position within the opening 276 by appropriate molded grips 282. A fitting feature 84 to control access to the opening 289 may be provided for the sample holder 278.

To complete the carrier 250, the end of the top member 258 may have an orifice 286 with downwardly extending flanges 288 adapted to hold a reaction vessel holder 90. The flanges 288 are concave on the inside to define a socket which compensates with the bulbous top on a reaction vessel 90 in a ball and socket joint manner. The lower portion of the reaction vessel holder 90 may be shaped as depicted in Figure 6 to have an inverted cavity or receptacle 92 at the upper end of which is hole 94 adapted to receive a pin 96 from a drive member, as will be described.

In an alternative embodiment of this invention, the reaction vessel holder 90 may be the reaction vessel itself although the use of the holder is preferred for its long term stability and reliability. If the reaction vessel 90 as a tube holder is adapted to receive a reaction vessel 100, the vessel has at the upper portion thereof a concentric chamber 102 for holding reaction reagents that typically may be used, for example, in an immunoassay process.

The reaction vessel holder 90 may be positioned in a thermal chamber 59 and is driven or nutated by an automatic 104 constructed in accordance with this invention and shown in detail in the cross sectional view of Figure 6. This drive apparatus mounted to the

bottom of the thermal chamber 59 and provides a
bidirectional motion as depicted by the line 306 (Fig.
9) as well as rotational motion as depicted by the line
108 to the reaction vessel holder 90. This drive
5 apparatus is powered by a single bidirectional drive
motor 110 which provides rotational motion to the drive
apparatus 104. The automatic apparatus engages the
reaction vessel holder 90 by elevating a mixing cylinder
or plate on which a the pin 96 is positioned contiguous
10 the periphery at a point off of the elongated axis of
the mixing plate. In other words the pin 96 engages the
bottom end of the mixing vessel 90 into a position which
is eccentric to the axis which mounts the mixing
cylinder 110. The apparatus then spins the plate moving
15 the engaged end of the vessel to an orbit. If the
vessel is managed so that it is free in two rotational
directions of freedom, then the contents of the reaction
vessel holder 90 will swirl or nutate thus mixing them.
Reversal of the drive which spins the mixing cylinder
20 110 stops the orbiting of the vessel and lowers the
cylinder thus disengaging the pin 96 from the reaction
vessel holder 90.

The drive 104 has a cylindrical housing or base 120
which has an internal flange 122 for mounting a bearing
25 124 which in turn mounts a nut 126. The lower end (in
the drawing) of the nut 126 is shaped to be cylindrical
and hollow to accommodate a screw 128 which is threaded
through the nut. At the top end of the screw is formed
an elongated cylindrical shaped mixer which defines a
30 mixing cylinder 110 which as described mounts the
eccentrically located pin 96. The lower end of the
screw 128 has a locknut or disk 130 engaged thereon to
limit the upper travel of the screw 128 in the nut 126.

Rotation of the mixing cylinder is prevented by a leaf spring type clamp 132, only a portion of which is shown, which engages the periphery of the mixing cylinder 110 to inhibit its motion up to a degree. The
5 leaf spring 132 is mounted to the housing 120. The lower portion of the nut 126 is shaped to have the form of a drive pulley 134 to accommodate a drive belt 131 from a motor 110 (Figure 2).

This automatic apparatus functions by rotating the
10 nut 126 with a drive belt attached to the pulley 134. The leaf spring 132 mounted on the base drags on the outside diameter of the cylindrical mixing cylinder 110 and thereby acts as a rotational clutch to the screw and mixing cylinder. When the nut is thus rotated, the
15 clutch prevents the screw from rotating so instead the screw elevates the mixing cylinder 110. This elevation continues until further elevation is prevented by the disk 130 at the bottom of the screw. At this point the clutch then slips allowing the nut 126, screw 128,
20 mixing cylinder 110 and disk 130 to rotate together. By this time, the pin 96 has risen (dashed line 133) to engage the recess and ultimately the bore 94 in the mixing vessel holder 90.

The engagement is complete when the screw reaches
25 the top of the travel and thereafter the eccentric motion of the pin 96 causes the bottom of the mixing vessel holder to rotate as depicted by the dashed line 134 and thereby causing vortexing to occur in the mixing vessel. If the rotation of the nut is reversed the
30 sequence starts over but in the opposite direction, i.e., the screw 128 is lowered until it strikes the flange 122.

This invention is seen to be very advantageous. It functions with a small number of inexpensive parts. One

drive motor permits the lifting and spin of the mixing cylinder. The device lends itself to be used with any number of duplicate devices and all to be driven by the same drive. It engages the bottom of the mixing vessel holder or mixing vessel gently so as to avoid on 5 spilling of the vessel contents. Furthermore the engagement of the pin 96 in the hole or bore 94 is a very positive, reliable drive even if one of the vessels is mispositioned.

10 The details of the rectilinear, incremental advance mechanism are depicted particularly in Figure 5. Thus in Figure 5 may be seen the platform 67 mounted at either end by linear slides 300. The linear slides are spring loaded by springs 302 to elevate the platform 67. 15 The linear slides 300 are driven by lever arms 304 which are pivoted at center pivot 306 and linked together with at the lower end 308 and adapted to act as a cam follower to be driven by a cam 310 driven by motors (not shown). The cam 310 is eccentric to raise and lower the 20 common point 308 and thereby raise and lower simultaneously the platform 67.

At the same time the platform 67 is connected to be moved laterally by a driven arm 312 which is actuated in turn by a thrust arm 314 which is tied to the cam at the 25 point 316 such that as a cam rotates the point 316 being at an eccentric point on the cam. The platform is moved laterally back and forth.

In operation the linear platform is driven by the motion of the thrust arm 314 acting on the lever arm 312 30 to be moved first to the right in the drawing to move a distance sufficient to move the carriers 12 from one position to the next position. Thereafter the action of the cam 310 on the common point 308 is such as to allow the platform to be lowered by the action of the springs

302 so as to remove the pins 18 from the bottom of the carriers 12. This point is depicted by the dash line 18' as described above. Next the platform 67 is moved to the left in the drawing as depicted by the arrow 75 so as to move the pins from a forward position to the next rearward position of the carriers 12. Finally the carriers again lifted as depicted by the arrow 69 and the pin motion 18' to engage the carriers of the next preceding position so that they too may be brought forward in the next cycle. The entire platform is mounted to ride upon support bars 320.

The several mixers 170 are driven by a belt 173 operated by a motor 172 driving pulleys 126. A single belt is used to drive the mixers 170 at the second, fourth and sixth positions in the incubation cycle. All of the first six processing positions are housed within a thermal housing 59. In addition, mixers 220 located at processing positions 8 and 9 are also driven by a belt 161 operated by a motor 160. A final mixer 192 is located also at position 10. It is operated by a drive motor 162 and belt 164. The thermal housing only encloses operating positions 1 through 7 as noted. A first translating unit 20 is a robotic system of conventional design and operates a probe with two degrees of freedom namely in the X and Z directions. The first translator 20 has a probe, which operates as will be described in Figure 12, to transfer sample from cap 80 into the mixing vessel 100 and reagent from the outer concentric portion 102 of the mixing vessel to the mixing vessel itself. In this connection, recall that the mixing vessel 100 is housed within a mixing vessel holder 90 which has the configuration described and is adapted to be nutated at its lower end.

At the wash positions 8 and 9, a wash means comprises a translating unit 410 (Figure 7) having a single Z motion direction and is adapted to drive a probe 400 having concentric 404 in order to aspirate the fluid from the reaction vessel 100 and introduce clean wash liquid. The needles are connected to a peristaltic pump 168 (Figure 12) through suitable tubing coaxial tubing 408. The translating mechanism designated by the numeral 410 is of a conventional type. At each wash position, a permanent magnet 412 which may be constructed of neodymium is attached to a lever arm 414 which is actuated at a pivot point 416 by a pulley drive 418. The pulley 422 is driven by a D.C. gear motor 421 to translate the magnet 412 near the reaction vessel holder 90 and then remove the magnet from the reaction vessel holder 90 as depicted by the dashed line drawing 424. This, as will be described, operates to attract the magnetic particles to the side of the reaction vessel so that the aspiration will not remove the magnetic carrier particles.

At the final or tenth processing position there is a second translating unit 32 of the same conventional design as those described heretofore and having two degrees of movement in the X and Z directions. This translator 32 is positioned at an angle such that processed sample fluid may be withdrawn from the reaction vessel 100 and moved over and injected into the fluid container 272 through its rubber septum 270.

A third transport 34 means seen most clearly in Figure 3 which is identical to and operates in the opposite sense to the first transport means 16 described previously. It too has folding pins (not shown) that operate to permit a pair of pins to be slid under the carrier 12 in that position, engage the receptacles in

the carrier 12 and then lock by being in the upright position to move the carrier from the processing chamber 19 to the inlet chamber 10. Once this has occurred, the movement of the transport means is reversed and the folding pins permitted to disengage from the bottom of the carrier and move back to engage another carrier in the processing chamber 19.

Finally a fifth transport means 36 (Figure 8) operates to move the carriers containing the completed sample reaction mixture into the exit area from which the process sample may be removed and transferred to an instrument for reading the results of the reaction. The liquid flow system is depicted in Figure 12. This system depicts how the various liquids are applied to the reaction vessel 100 at the several positions to the 1, 8, 9 and 10 in the processing chamber. The first position is not shown. It is recalled that in this position sample and reagent and water each separated by an air bubble are withdrawn from the carrier and water supply 450 and applied to the reaction vessel at the first position. After this reaction vessel are processed through the processing chamber.

The liquids are processed by using a syringe type pump 452 (Fig. 12) which is motor driven and actuated by the process controller 500. A peristaltic pump 168 is (1) to withdraw fluid from the wash buffer 458 and (2) provided to supply waste fluid to a waste disposal 460. The syringe type pump 452 operating through a rotary valve 454 supplies fluid from the resuspension buffer supply 464, a source of water 450, and the probe wash supply 466 for supply to the translators 20 and 32.

In operation, after having received the sample reagent and water at the first processing position the reaction mixture is allowed to incubate at positions 2

through 6. It is as previously described, vortexed position 2, 4 and 6. Following incubation, the next two units 8 and 9 are in the wash chamber 26. In this chamber, the magnets are applied to the side of the reaction vessel as previously described to cause the magnetic particles to clump at the sides. This allows the contents of the reaction vessels at both positions 8 and 9 to initially have the liquid withdrawn therefrom and passed to the waste bottle 460. Thereafter fresh wash buffer is introduced from 458.

After passing through the wash station, the carrier arrives at position 10. At this position the reaction vessel is allowed to arrive virtually dry so that suspension buffer 464 may be added to the reaction vessel. The probe is washed thereafter with a suitable wash solution from the supply 466 such as sodium hypochloride plus sodium hydroxide. The mixer at this position disperses the magnetic particles in the buffer. After this, the resuspended buffer is drawn up into the translator 32. The translator is repositioned and the withdrawn contents of the reaction vessel inserted into the container 269. Following this action the carrier is moved back into the inlet chamber.

The mechanism for bringing this back to the inlet chamber is substantially the same as the transport mechanism which brought the carrier from the inlet chamber to the processing chamber except that it operates in reverse as described. This is seen partially in Figure 8. As depicted in Figure 8, the carrier 12 is driven by the pin 400 from the carrier transport 402 into the inlet chamber. At this point the carrier 402 reverses and returns to the processing chamber. A lever arm 404 is mounted on a slider mechanism 406 and is actuated by a wire 408 wrapped

around a pulley 410. The other end of the wire 408 is attached to the carrier drive 402 such that as it returns to the processing chamber the wire is tightened causing the lever arm 404 to be drawn against the carrier 12 shifting it to the position 412 illustrated by the dashed line. A spring 414 is attached to the slider arm 406 to return it and the lever 404 back to a withdrawn position when the carrier drive 402 again returns to the inlet area.

10 The apparatus utilizes a single time-template for processing the immunochemistry. The operations performed on each assay follows a fixed sequence and is governed by a fixed timing cycle. The operation sequence begins at position #1 (Figure 1) where the patient sample, reagent and magnetic particles are mixed together to initiate the assay. Positions #2 through #7 are the incubation stations. Assays passing through these stations would received 6 cycles worth of incubation at the set temperature of 37°C. As the assays proceed to position #8 and #9, the dual wash stations would provide a total of three washes. The consumables are left "dry" as they advance into the exit station at position #10. At position #10, the captured analytes on the magnetic particles are resuspended and then injected into the aca® pack in the carrier.

Because the assays are to be run in an assembly line style, the stations are separated by their tasks and are programmed to function independently. A set of flowcharts are provided in Figure 13A-13G to illustrate the processing structure of the instrument. These flowcharts are the basis of programming the CPU.

Description of Flow Charts

Figure 13A gives the overview of processing. The five tasks shown: method decode task, entrance and exit stations task, wash stations task, incubation task and the transport task are initiated upon startup. Once started up, these tasks are expected to run as independent entities. Along with initiating the tasks, a method processing queue and task status flags are required in memory.

At the start of the processing loop in Figure 13A, the input tray area 10 and the processing chamber (positions #1-#10) are checked as to whether carriers containing tests are in the instrument. The input tray has sensors (not shown) to detect carriers 12 waiting for processing. Once the carriers 12 begin their cycle in the processing chamber, the method processing queue keeps track of their locations. If any carrier is pending for processing or is undergoing processing, the four tasks: method decode, entrance and exit stations, wash stations and incubation, should check whether any carrier location are in their domain for processing. If there is, then the task should raise a busy status, proceed with processing, and clear their status flag when done. Once all the tasks are given the signal to begin, the processing loop would delay for the duration of one full cycle time. This operation is shown on Figure 13A. The dotted line indicates the signal flow between the main processing loop to the different tasks. After the full cycle delay, the loop would continue by checking the status flag of each task. If all tasks are done, then the transport task is initiated to advance the carriers to their next position and the carrier locations are updated in the method processing queue.

(Figure 13G) However, if there are any busy tasks, then the system would raise a cycle time error and delay for a second prior to rechecking the status of all the tasks. After advancing the carriers to their next positions, the main processing loop would start again from the beginning until all carriers are processed.

Figure 13B gives the flowchart for the method decode task. If there is a carrier pending in the input tray, the method decode task would raise its busy flag. The aca® pack 78 is decoded to determine which method type that carrier 12 contains. From the method type, the system would obtain from memory storage, the method specific parameters. This information is placed into the method processing queue and the carrier position for that test is set to 0. At this point, the carrier 12 would wait for the rectilinear transport (Figure 5) to advance the carrier locations in the method processing queue and the entrance task to shuttle it into processing chamber 18. The method decode task's busy status is reset. And the method decode task ends.

Figures 13C and D gives the flowchart for the entrance and exit stations. Since the entrance and exit station shares the same precision fluid delivery pump, they require timing coordination to avoid conflicts in resource usage. Because of this, the processing cycle is further subdivided into two halves. The first half of the cycle is allotted to the exit station processing; while the second half belongs to the entrance station. When this task receives the start signal, it would check the method processing queue for the presence of a carrier 12. If any carrier location in the queue = 1, then the carrier 12 in decode area 14 would be shuttled into position #1 where a possible preheating unit would

bring the reaction vessel to approach the incubation chamber temperature.

In case the exit station may not finish processing in the allotted half cycle time, an entrance standby flag, set and reset by the exit station, is monitored after the entrance station delay to insure against any resource conflicts. After the delay to the 2nd half of the cycle and the standby flag indicating the pump resource is free, the entrance arm would proceed to aspirate the method specific volume of fluid from the aca® sample cup on the carrier, aspirate the method specific volume of reagent from the consumable collar well and dispense both into the reaction vessel center well. The entrance arm needle is washed at this point. The entrance station busy flag is reset. While station #1 is running, if a carrier exists at position #10, the exit station would raise its busy flag and set the entrance station standby flag as well (Fig. 13D). This standby flag would give the entrance station an indication that the pump resource is busy. After the flags are set, the exit station needle would deliver a method specific volume of resuspension buffer into the center well of the reaction vessel. The mixer (Figure 6) would spin up in one direction to resuspend the magnetic particles. The mixer would then be retracted by spinning in the opposite direction. The exit station needle would pick up a method specific amount of magnetic suspension and dispense it along with a method specific volume of resuspension buffer chase into the aca® pack 269. The instrument would then wash the needle. After the needle washing routine is completed, the entrance standby flag may be reset to 0. This would allow the entrance station to begin its processing. At this point, the carrier 12 is free to be shuttled out of

the processing chamber 18. Once the carrier 12 is out, the exit station task is completed. If both the entrance and exit station's status flag indicated that they are done, then the entrance/exit task is completed and should exit.

Figure 13E shows the flowchart of the dual wash stations at position #8 and #9. When the wash station task receives the start signal from the main processing loop, it checks the method processing queue whether position #8 or #9 is filled. If a carrier 12 is in either station, then the wash station task raises a busy flag. Each wash station is required to go through two aspirate-dispense cycles in order to provide three complete clean washes. So the first operation is to set a loop variable to 1. The magnets are then moved against the consumable to begin the magnetic particle separation process. After waiting a number of seconds to allow the magnetic field to pull the magnetic particles into a pellet against the reaction vessel wall, the wash station task starts the peristaltic pump to pull out the "dirty" fluid. The valves for the aspirate lines at position #8 and #9 are opened and the wash probes are moved into the consumables. After allowing several seconds for the pump to suck the consumables dry, station #8 and #9 are checked for presence of an assay so as to determine which dispense lines may be opened along with the aspirate lines to allow rinsing of the probe. For station #9, it has an extra requirement that it should be the first wash loop before the dispense line would open. This is because the reaction vessels need to come out of the second wash station dry so that a resuspension buffer can be added at the exit station, position #10. After the dispense lines(s) are opened for a set amount of rinsing time,

the aspirate line valves are closed. The dispense line(s) would remain open to dispense the specified amount of new wash buffer. Then the dispense valve(s) are closed, the pump is stopped, and the magnets and probes are withdrawn. The wash stations mixers are spun to its up position to resuspend magnetic particles in the new wash buffer. The mixers are then spun in the opposite direction to retract them. This completes the loop. The loop counter is incremented. The wash sequence is started again for the second aspirate-dispense cycle. After the second cycle is completed, the wash station task is done and it should exit.

Figure 13 F shows the flowchart of the incubation task. Upon receiving the start signal from the main processing loop, it checks the method processing queue for carriers in positions #2, #4, and #6. If a carrier is in any of those positions, the incubation stations busy flag is set, and the incubation mixer spins for 20 seconds to keep the chrome particles dispersed in the sample and conjugate reagent mixture. At position #2, this mixing is also required to aid the dissolution of any tablets in the consumable center well. After 20 seconds, the mixers would spin in the opposite direction to retract them back to the down positions. Since the incubation temperature is controlled by electronic hardware, the incubation task does not have to be concerned about temperature control. At this point, the incubation task may reset the busy flag and should exit.

In the Claims:

1. An automatic multilinear apparatus for processing immunoassays of samples using a solid support, the assays having bound and free phases, the bound phase being bound to the solid support, the apparatus comprising:
- an inlet chamber,
 - a plurality of carriers positioned in the inlet chamber each carrier holding a sample, a rotatably mounted reaction vessel, particles responsive to a magnetic field, reagents, and a reaction product container,
 - a processing chamber generally parallel to the inlet chamber,
 - first means to transport the carriers linearly in a first direction to one end of the inlet chamber,
 - second means to transport sequentially the carriers linearly in a second direction transverse to the first direction from the one end of the inlet chamber to one end of the processing chamber,
 - first translating means for acting on each carrier at the one end of the processing chamber to transfer each carrier's sample and reagents into the carrier's reaction vessel,
 - third means to transport sequentially the carriers linearly in a third direction opposite to and generally parallel to the first direction to several processing positions,
 - means for vortexing at at least one processing position by nutating the lower portion of each carrier reaction vessel,
 - wash means at at least one processing position for removing liquids from each carrier's reaction vessel and replacing the liquids with a different liquid,

magnet means at each wash means position for applying a magnetic field to each carrier reaction vessel prior to liquid removal,

second translating means at the last
5 sequential processing position for transferring the contents of each carrier's reaction vessel to its container,

fourth means to transport each carrier transversely of the third direction from the processing
10 chamber to the other end of the inlet chamber, and

fifth means to transport each carrier from the other end of the inlet chamber in a direction generally parallel to the first direction for storage.

2. The apparatus of Claim 1 wherein each carrier
15 provided with at least a pair of receptacles positioned at the bottom of the carrier, and the second means has a pair of spring loaded pins foldable downwardly in the second direction, whereby the second means can be displaced in a direction opposite the second direction
20 to engage each pair of receptacles and transport each carrier in the second direction.

3. The apparatus of Claim 1 wherein the carrier defines a mounting orifice lined by a flexible material, the reaction vessel top portion being positioned in the
25 orifice so that the lower end is nutatable.

4. The apparatus of Claim 1 which also includes a thermal chamber adapted to substantially enclose the processing positions.

5. The apparatus of Claim 1 wherein the third
30 means includes pairs of fixed pins adapted to engage each of the plural carrier receptacles, and rectilinear transport means for raising each of the pair of fixed pins to engage the receptacles and displace the

carrier's one processing position, and lowering each the pair of fixed pins to disengage the receptacles.

6. The apparatus of Claim 1 wherein the magnet means includes a magnet, and means to displace the magnet to engage the side of a reaction vessel, thereby to apply the magnetic field to the particles.

7. The apparatus of Claim 1 wherein the carrier defines a mounting orifice lined by a flexible material, the reaction vessel top portion being positioned in the orifice so that the lower end is nutatable.

8. The apparatus of Claim 1 which also includes a thermal chamber adapted to substantially enclose the processing positions.

9. The apparatus of Claim 1 wherein each carrier is provided with at least a pair of receptacles positioned at the bottom of the carrier, and the second means has a pair of spring loaded pins foldable downwardly in the second direction, whereby the second means can be displaced in a direction opposite the second direction to engage each pair of receptacles and transport each carrier in the second direction.

10. The apparatus of Claim 1 which also includes a thermal chamber adapted to substantially enclose the processing positions.

11. The apparatus of Claim 1 wherein the third means includes pairs of fixed pins adapted to engage each of the plural carrier receptacles, and rectilinear transport means for raising each of the pairs of fixed pins to engage the receptacles and displace the carriers one processing position, and lowering each pair of fixed pins to disengage the receptacles.

12. The apparatus of Claim 1 wherein the third means includes pairs of fixed pins adapted to engage each of the plural carrier receptacles, and rectilinear

transport means for raising each of the pairs of fixed pins to engage the receptacles and displace the carriers one processing position, and lowering each pair of fixed pins to disengage the receptacles.

5 13. The apparatus of Claim 1 which also includes a thermal chamber adapted to substantially enclose the processing positions.

14. The apparatus of Claim 1 wherein the magnet means includes a magnet, and means to displace the
10 magnet to engage the side of a reaction vessel, thereby to apply the magnetic field to the particles.

15 15. The apparatus of Claim 1 wherein the magnet means includes a magnet, and means to displace the magnet to engage the side of a reaction vessel, thereby to apply the magnetic field to the particles.

16. The apparatus of Claim 1 wherein the third means includes pairs of fixed pins adapted to engage each of the plural carrier receptacles, and rectilinear
20 pins to engage the receptacles and displace the carriers one processing position, and lowering each pair of fixed pins to disengage the receptacles.

17. The apparatus of Claim 1 wherein each carrier is provided with at least a pair of receptacles
25 positioned at the bottom of the carrier, and the second means has a pair of spring loaded pins foldable downwardly in the second direction, whereby the second means can be displaced in a direction opposite the second direction to engage each pair of receptacles and
30 transport each carrier in the second direction.

18. The apparatus of Claim 1 wherein the fourth means has a pair of spring loaded pins foldable downwardly in the third direction, whereby the second means can be displaced in a direction opposite the third

direction to engage each pair of carrier's receptacle to transport such carrier.

19. The apparatus of Claim 1 wherein each carrier is provided with at least a pair of receptacles
5 positioned at the bottom of the carrier, and the second means has a pair of spring loaded pins foldable downwardly in the second direction, whereby the second means can be displaced in a direction opposite the second direction to engage each pair of receptacles and
10 transport each carrier in the second direction.

20. The apparatus of Claim 1 which also includes a thermal chamber adapted to substantially enclose the processing positions.

21. The apparatus of Claim 1 wherein the third
15 means includes pairs of fixed pins adapted to engage each of the plural carrier receptacles, and rectilinear transport means for raising each of the pairs of fixed pins to engage the receptacles and displace the carriers one processing position, and lowering each pair of fixed
20 pins to disengage the receptacles.

Fig. 1

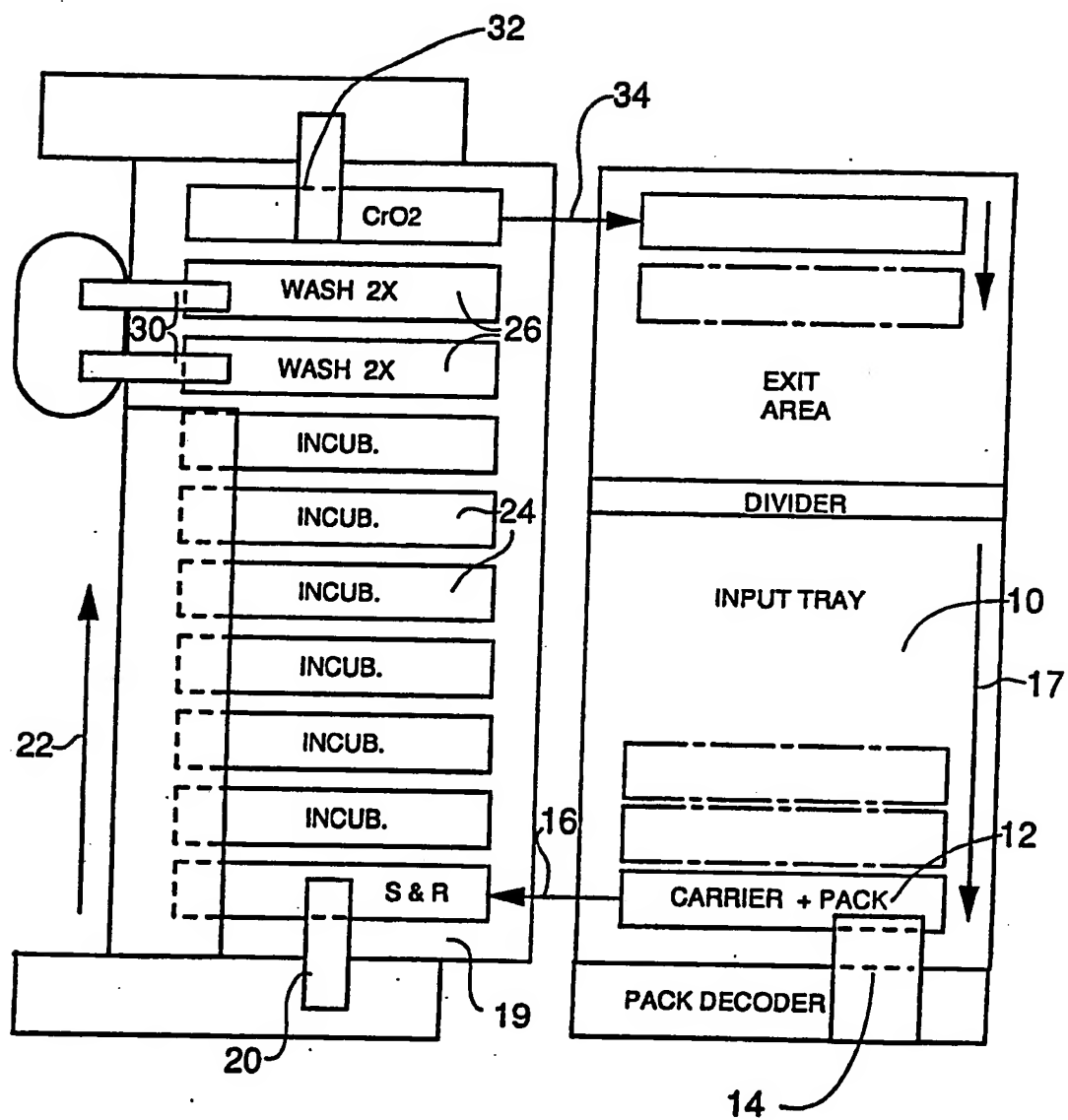
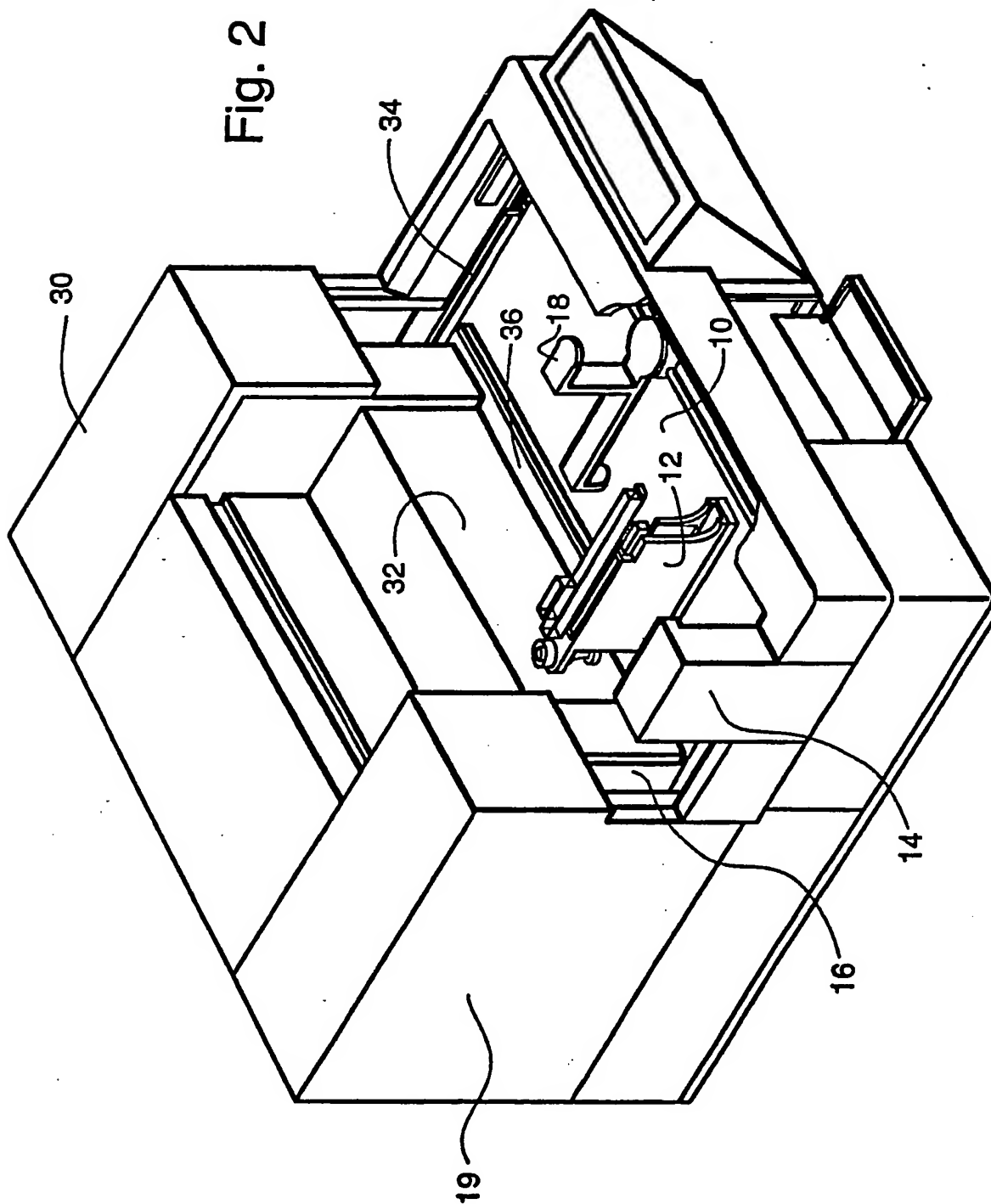


Fig. 2



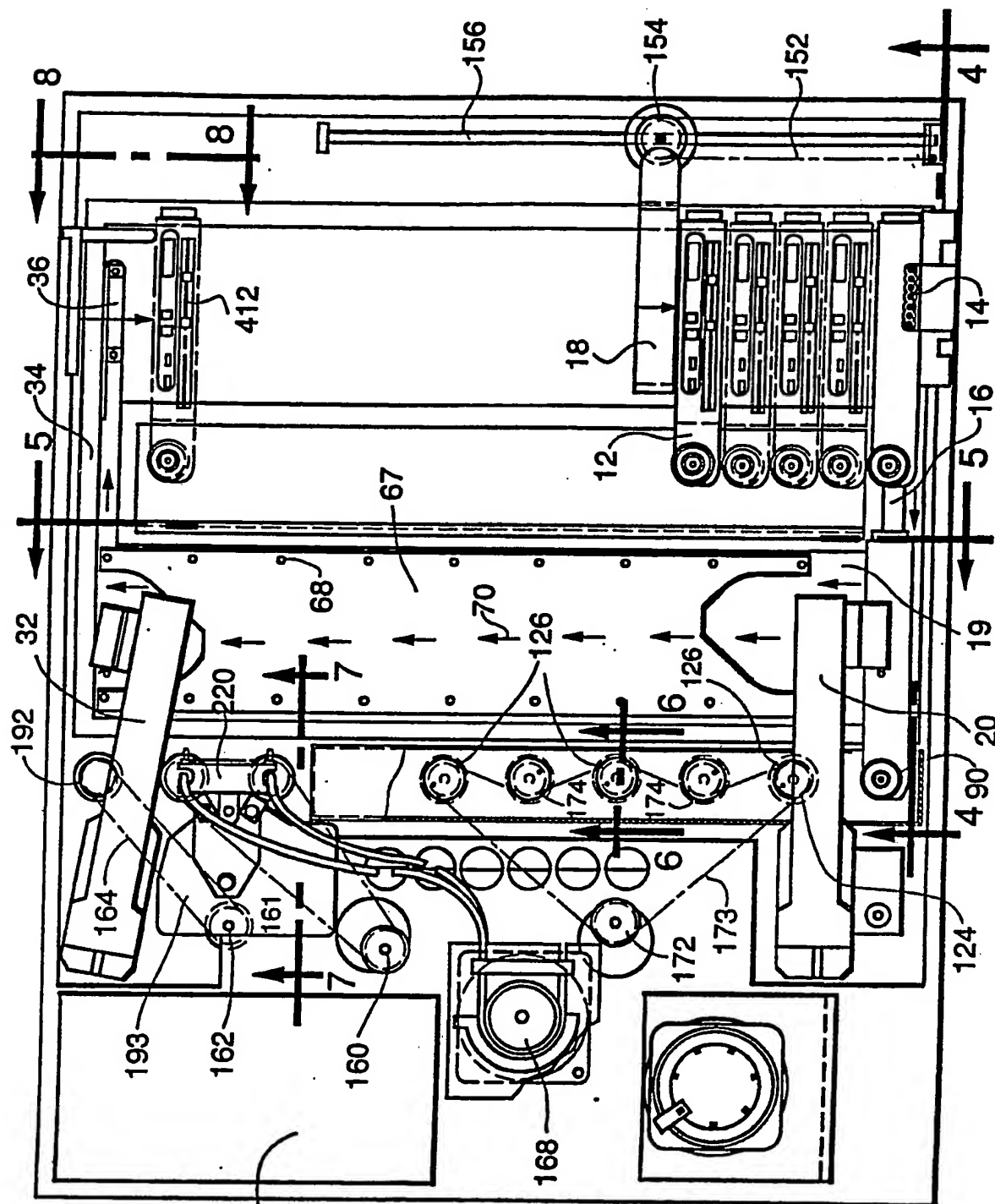


Fig. 3

500

Fig. 4

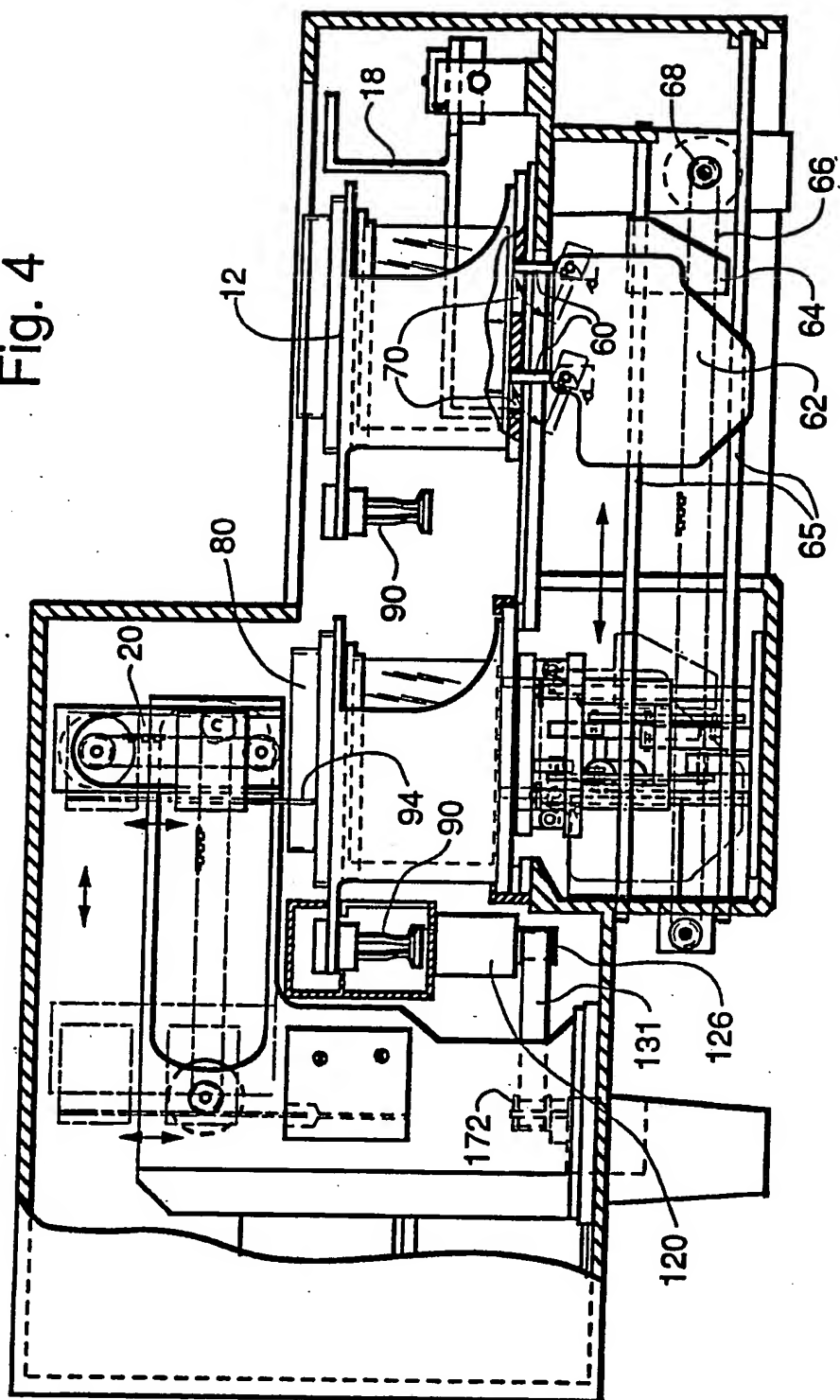


Fig. 5

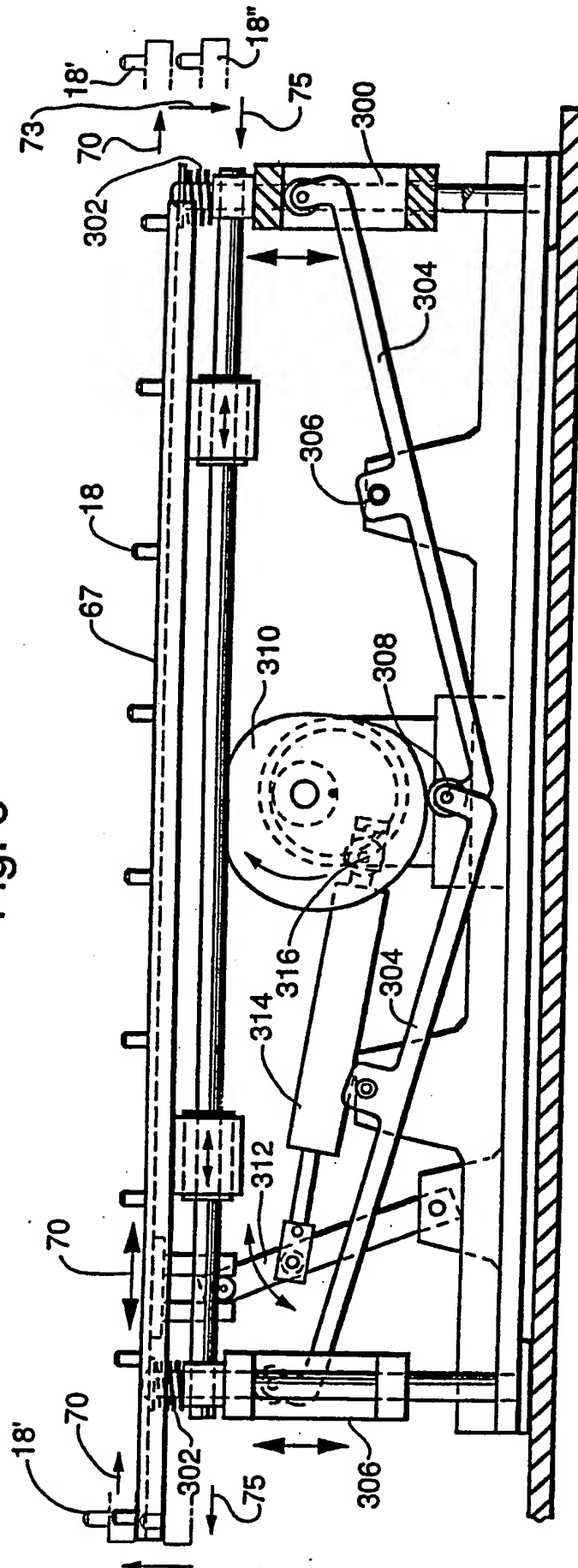
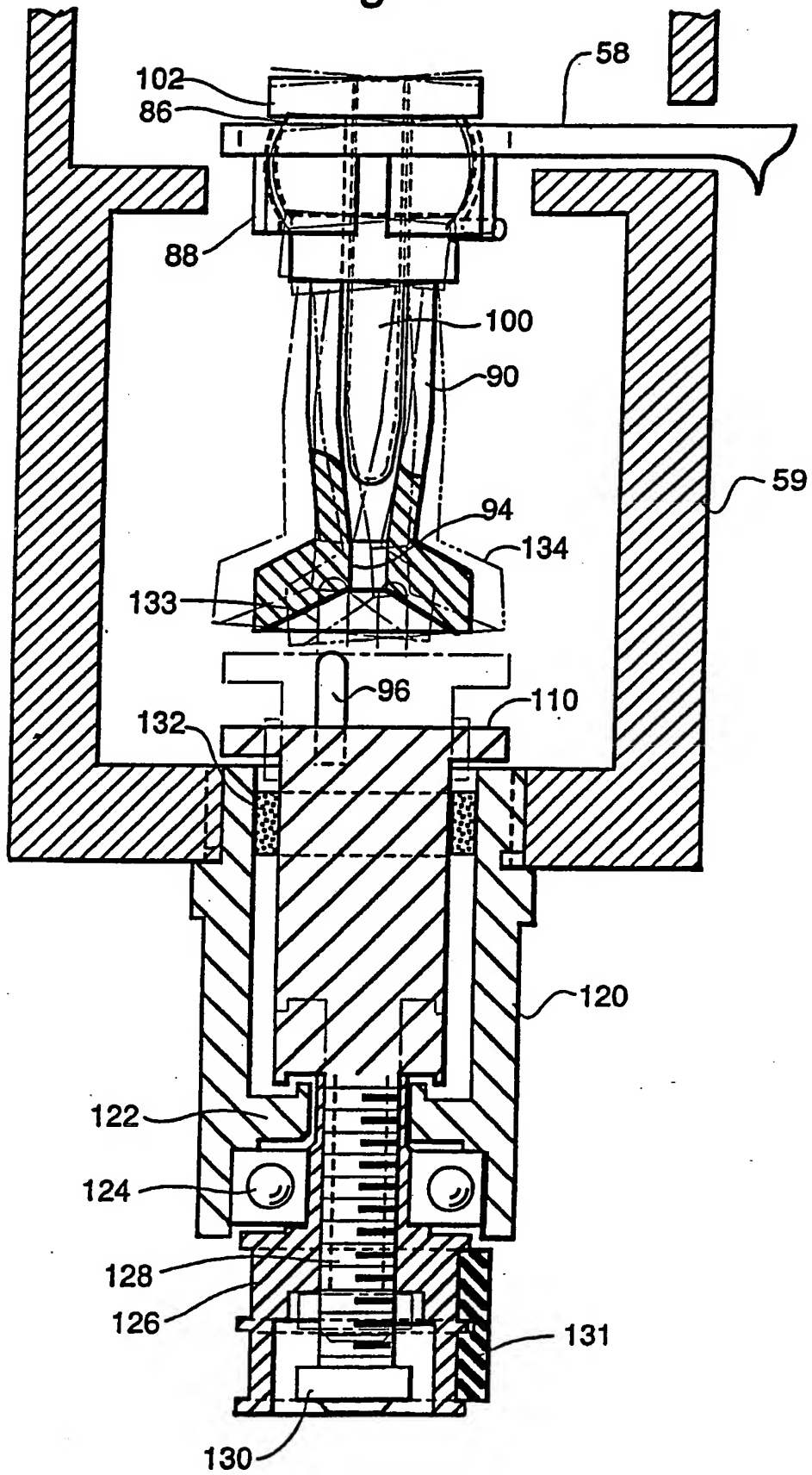
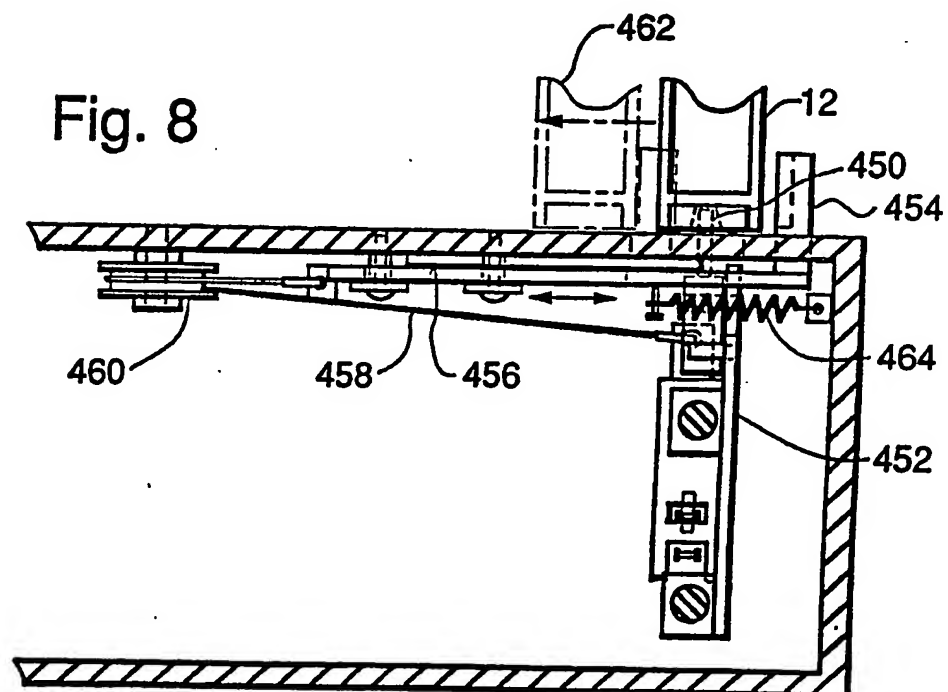
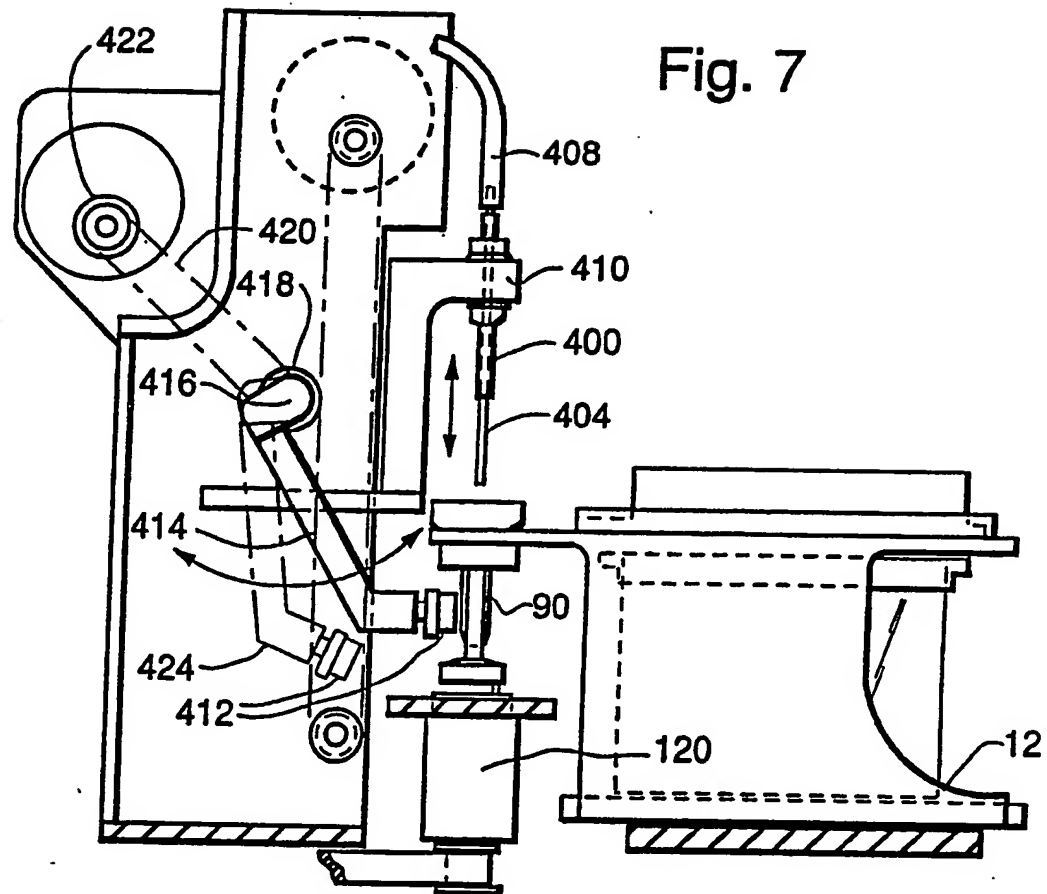


Fig. 6





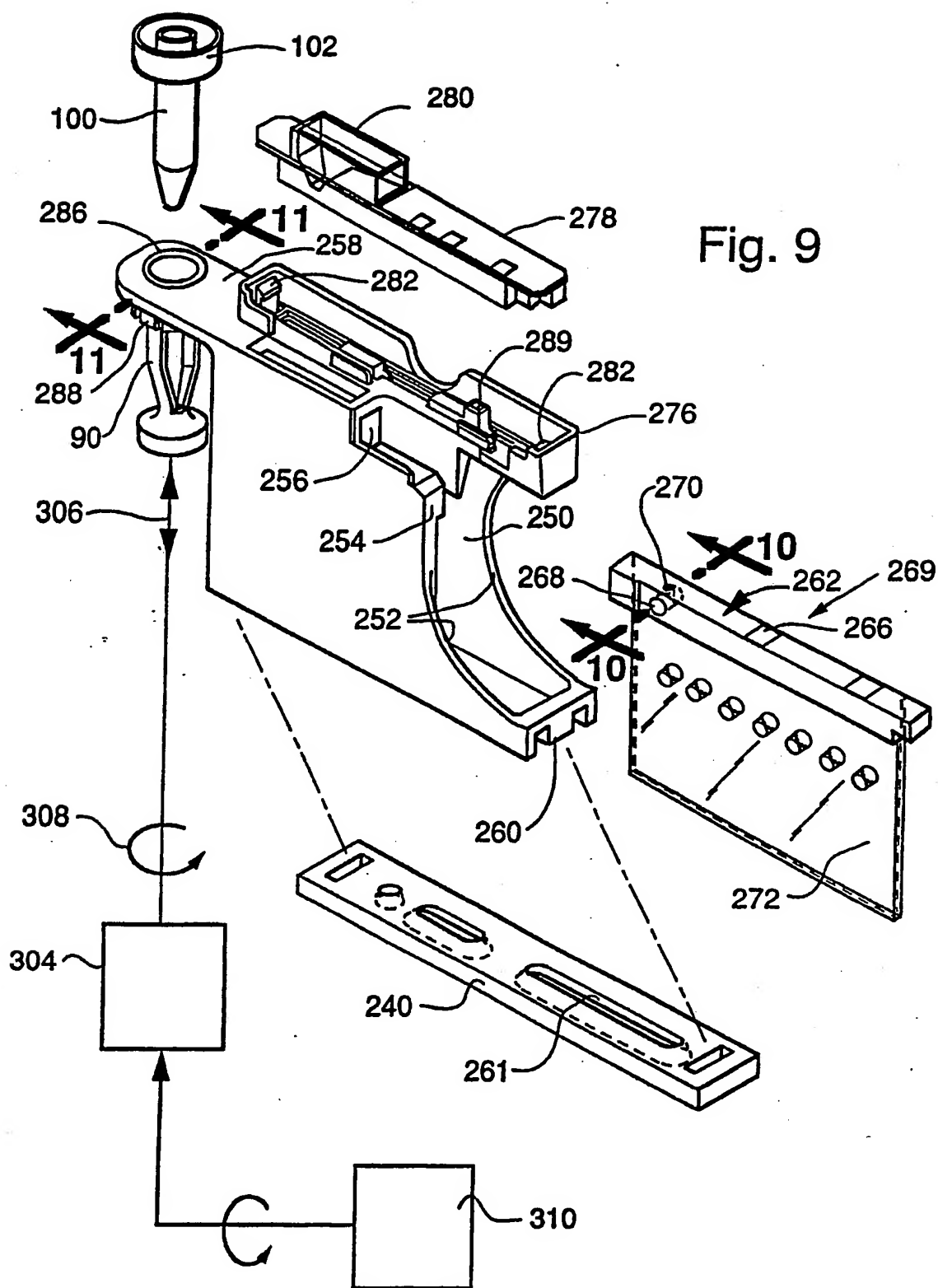


Fig. 10

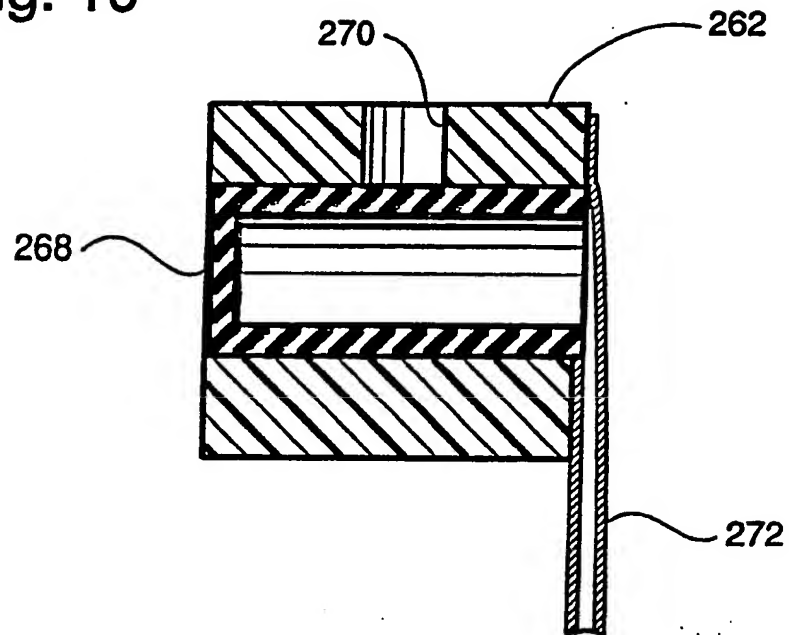


Fig. 11

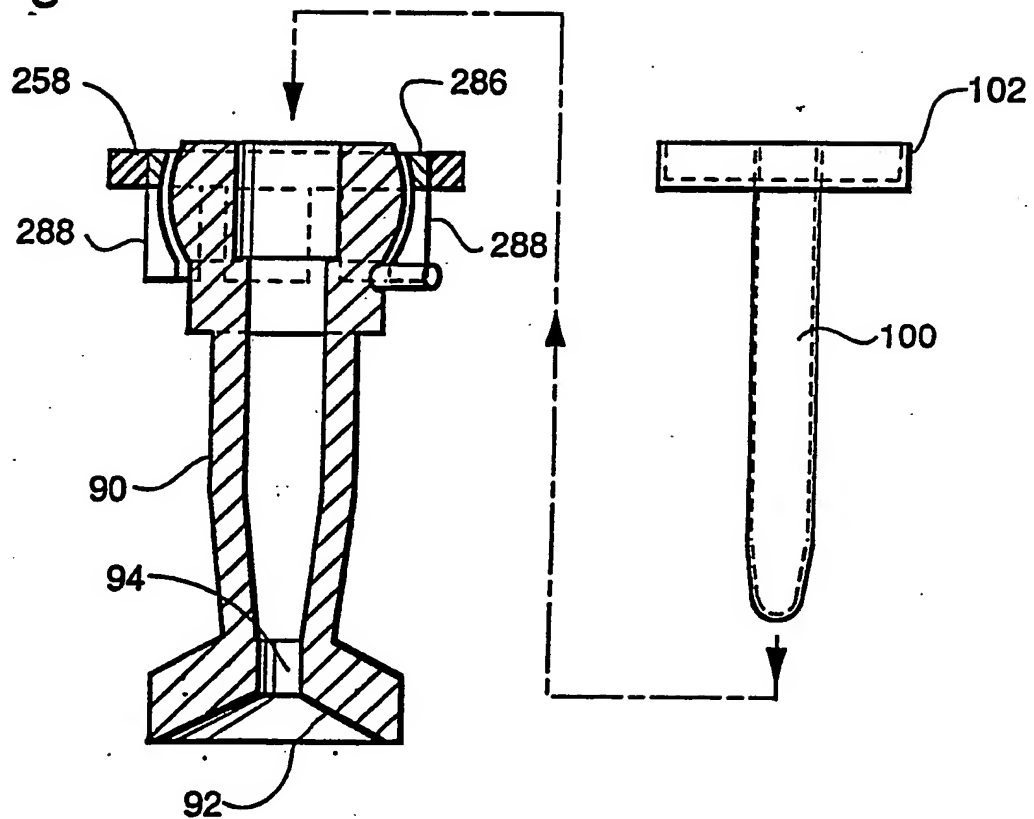


Fig. 12

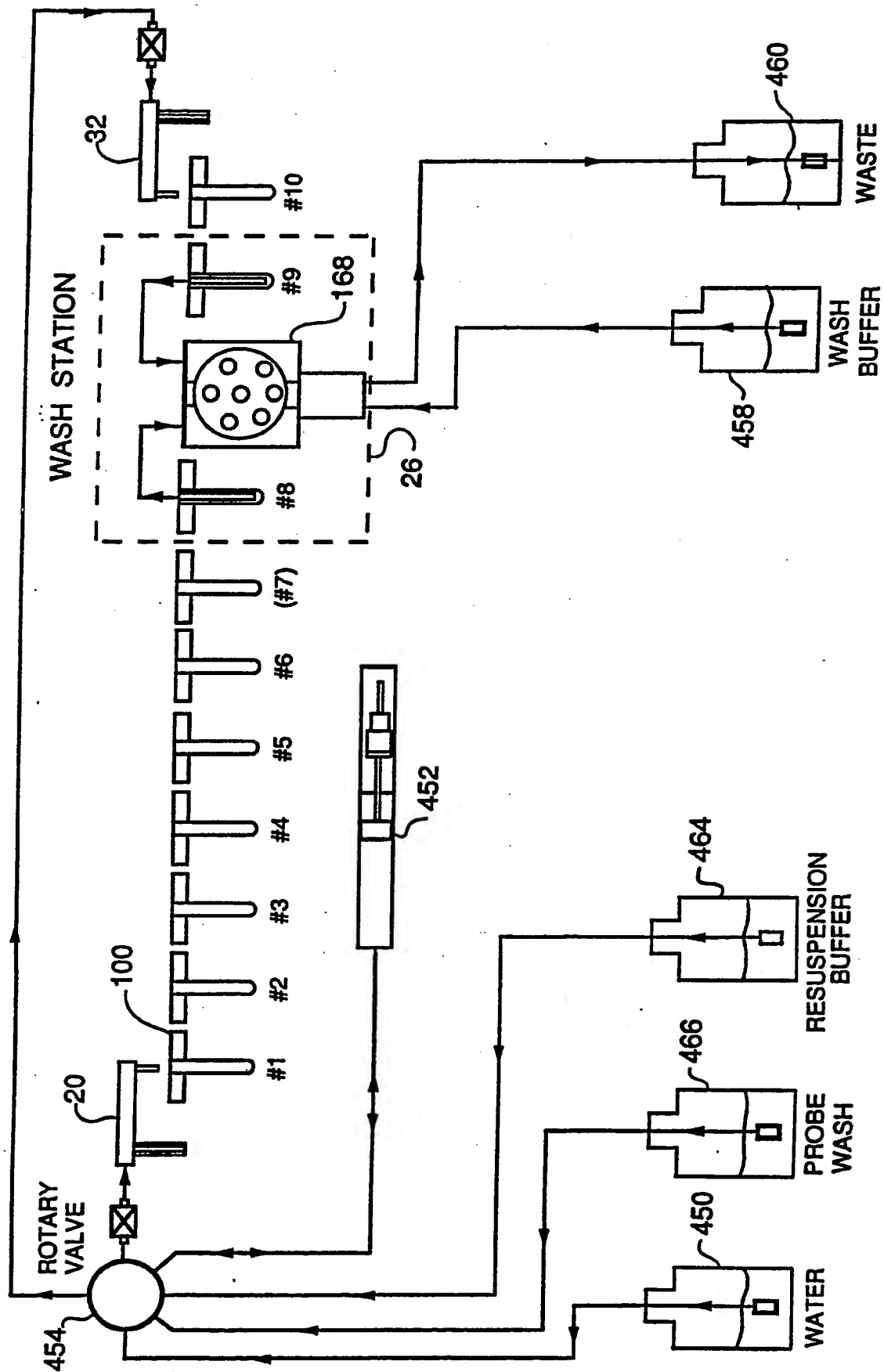


Fig. 13A

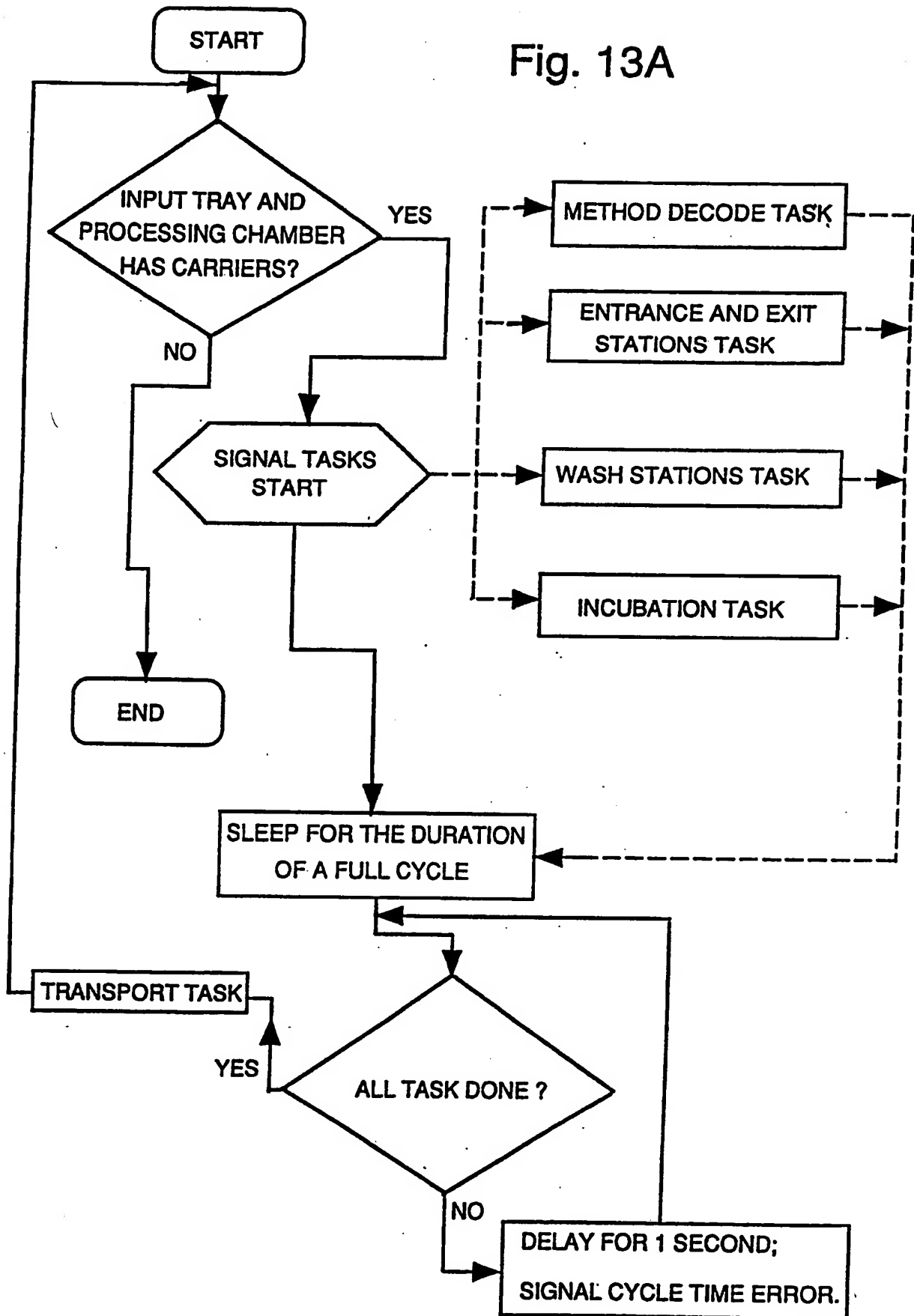


Fig. 13B

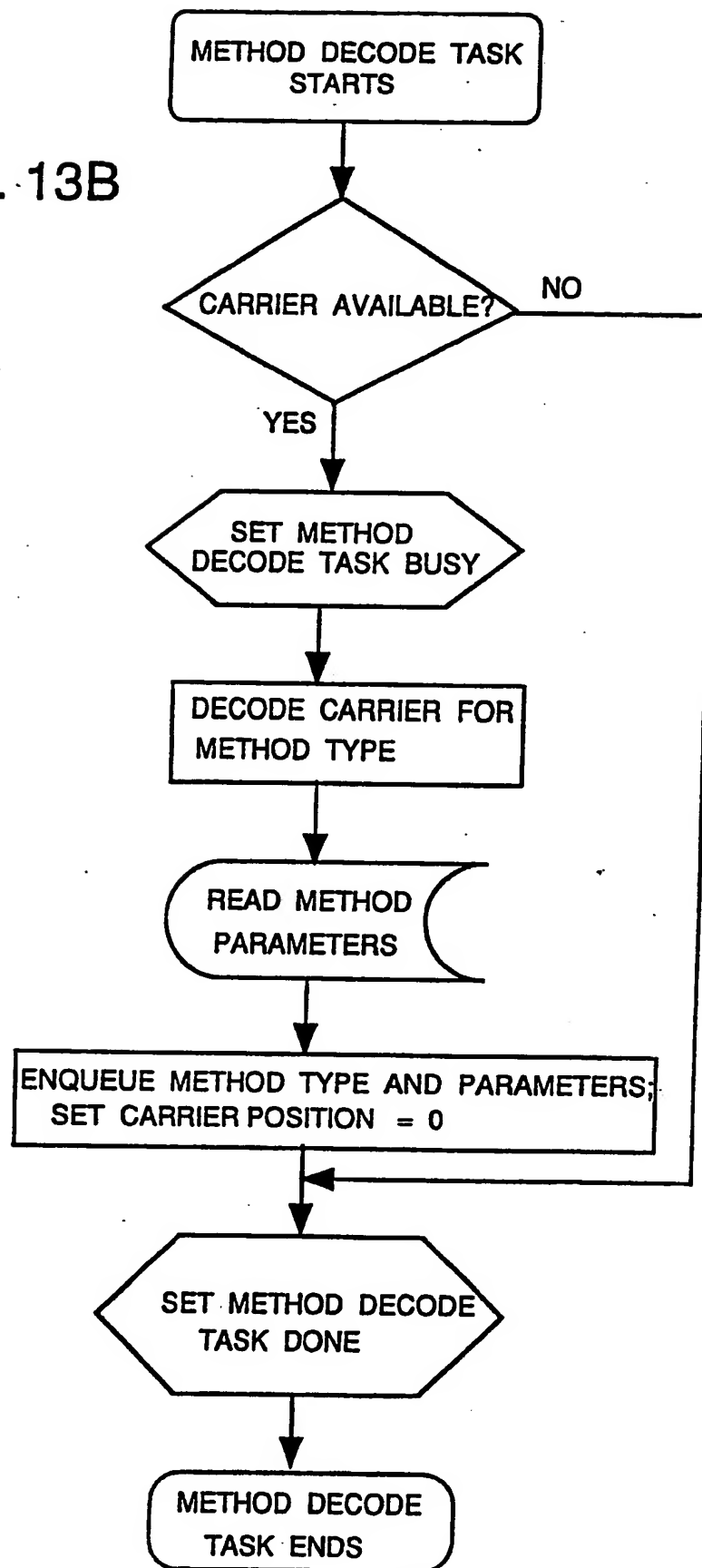


Fig. 13C

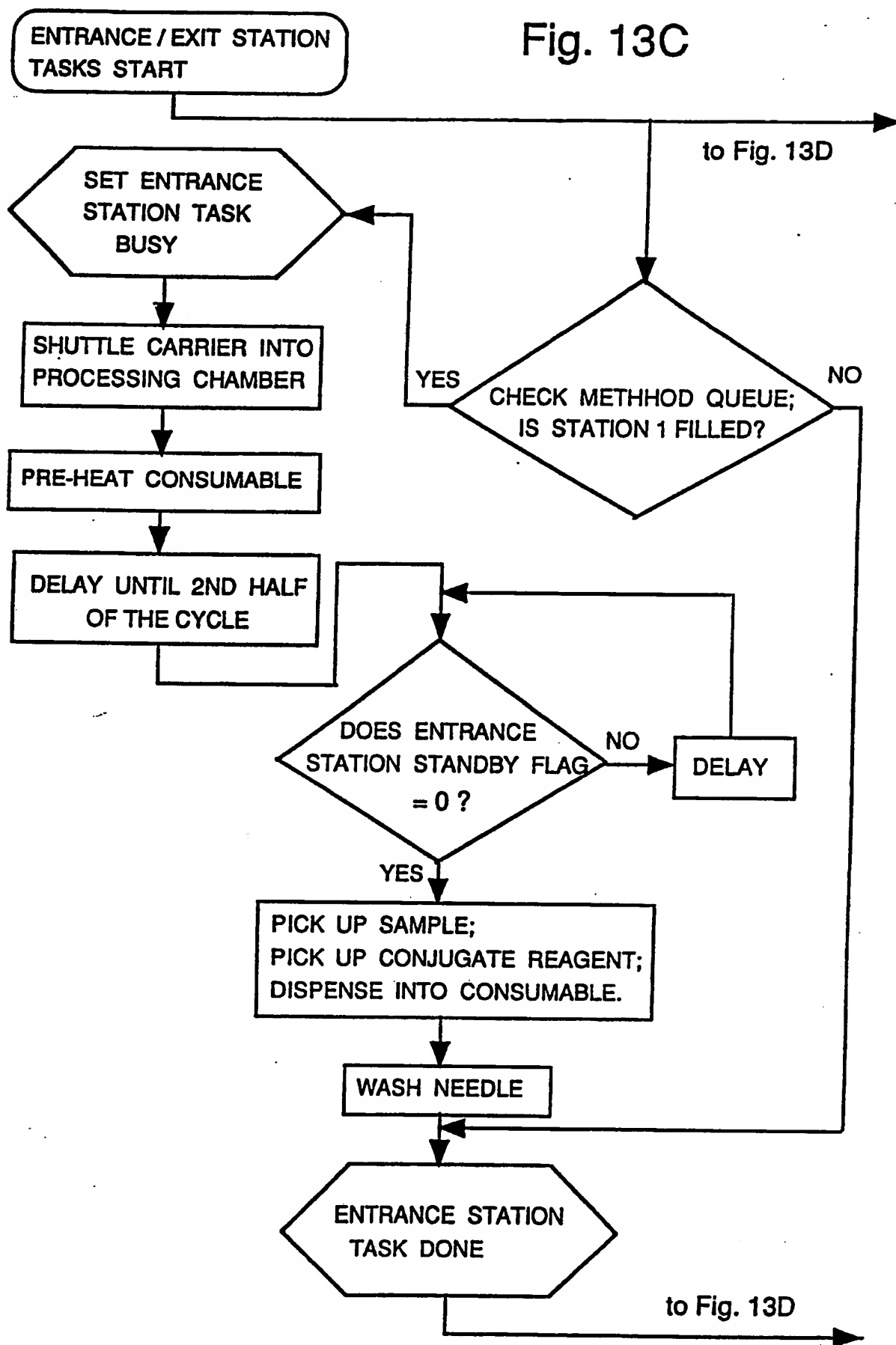


Fig. 13D

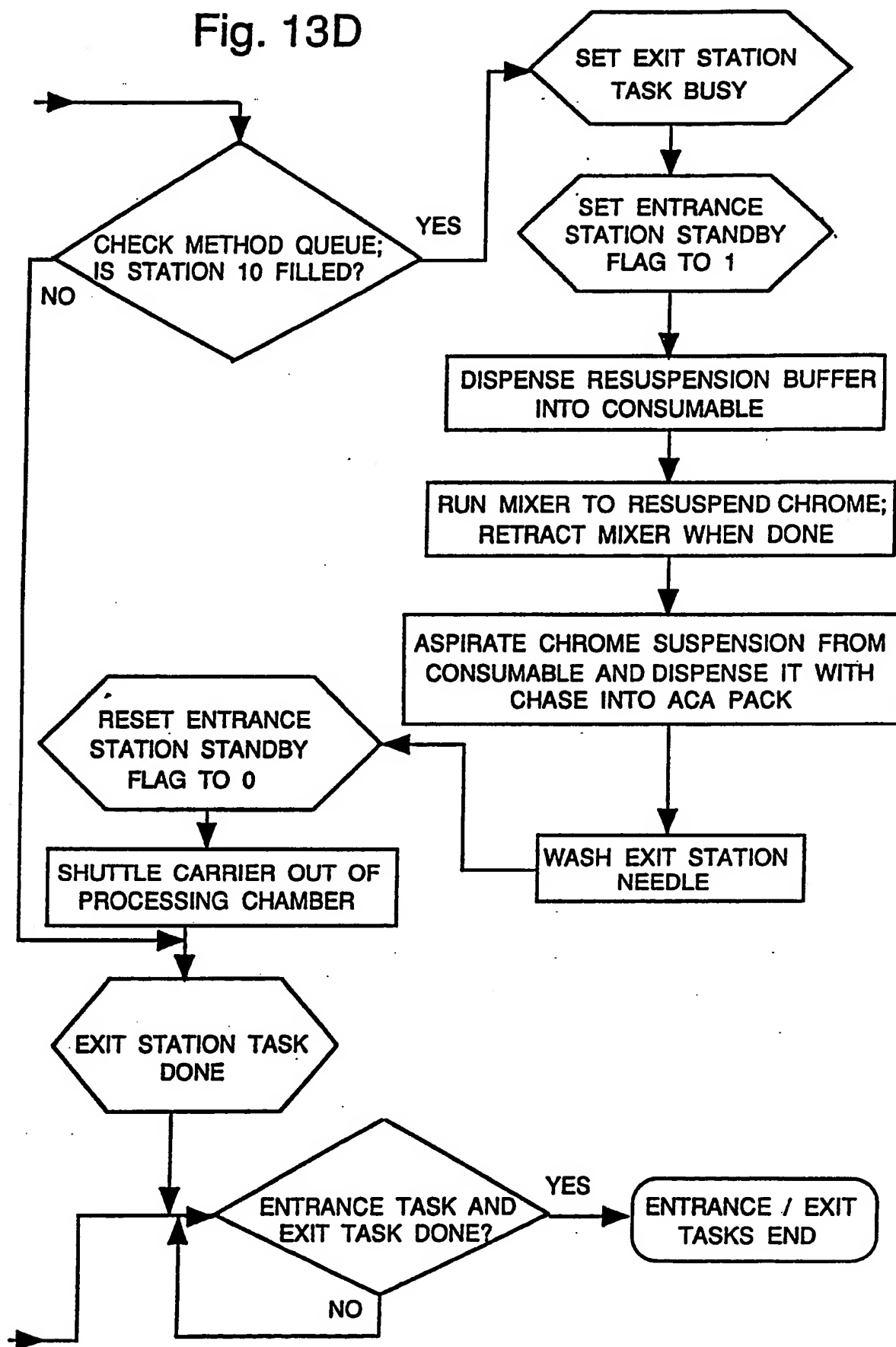


Fig. 13E

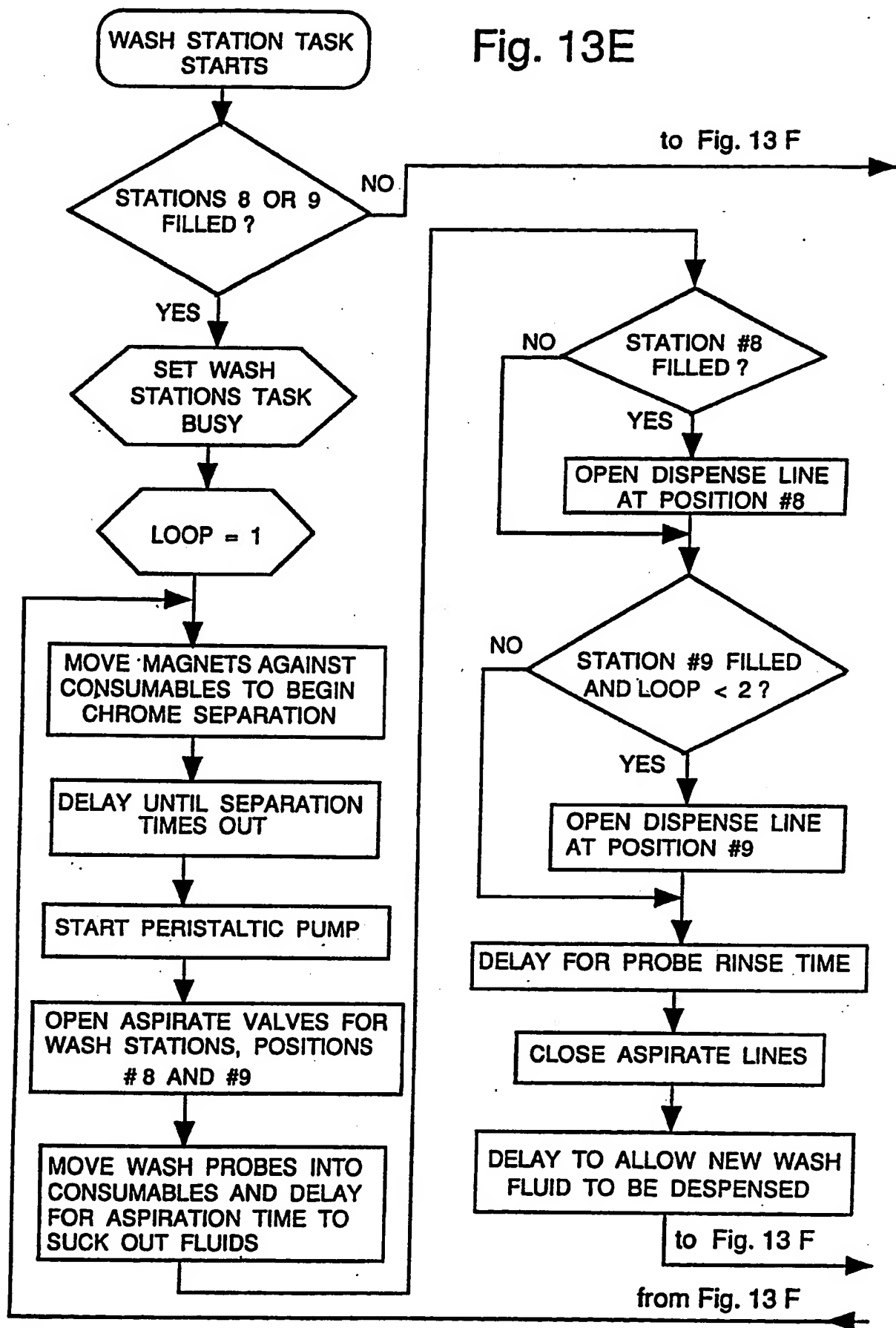


Fig. 13F

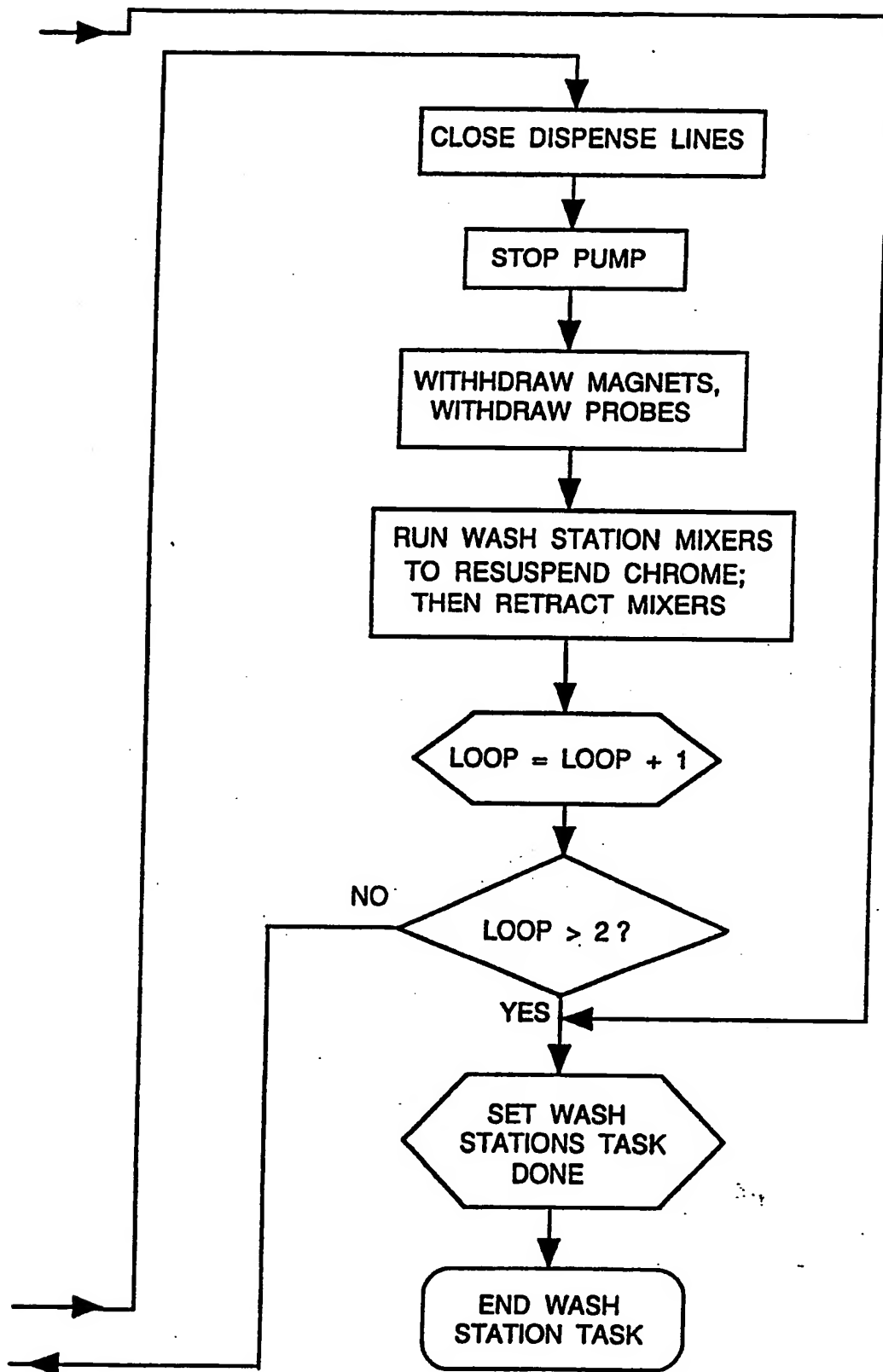


Fig. 13G

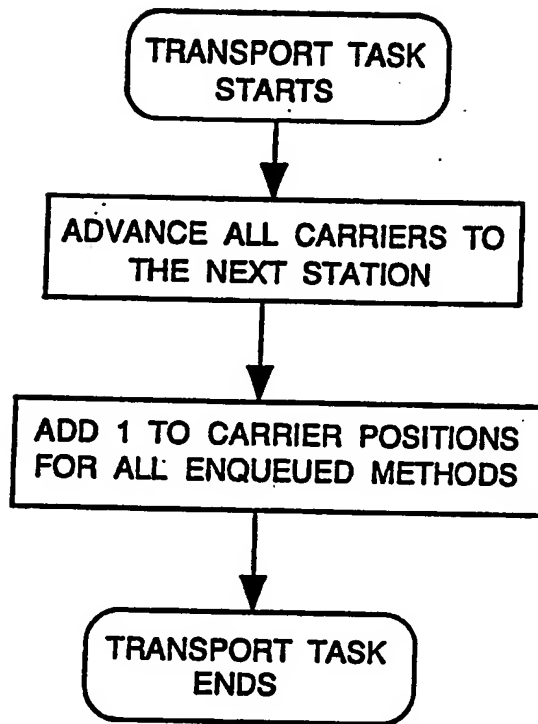
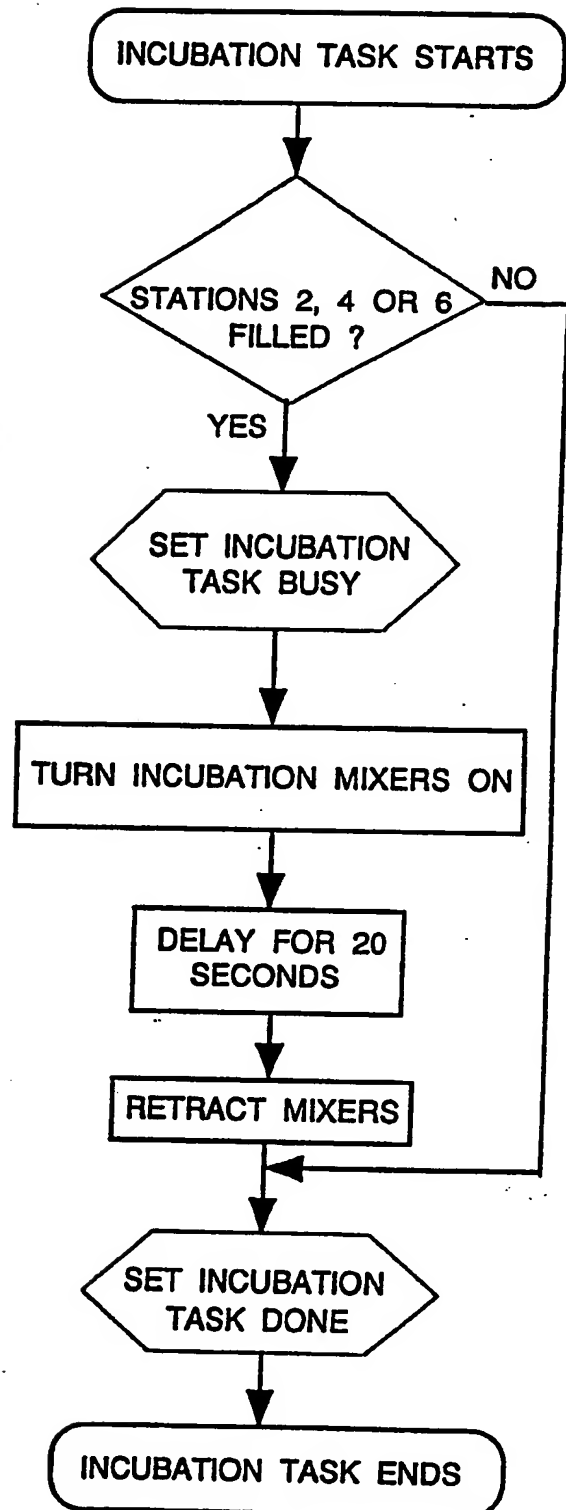


Fig. 13H



INTERNATIONAL SEARCH REPORT

International Application No PCT/US 92/06058

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all) ⁶ According to International Patent Classification (IPC) or to both National Classification and IPC IPC5: G 01 N 35/02						
II. FIELDS SEARCHED <div style="text-align: center; border-top: 1px solid black; border-bottom: 1px solid black;">Minimum Documentation Searched⁷</div> <table style="width: 100%; border-collapse: collapse;"> <tr> <td style="width: 20%; border-bottom: 1px solid black;">Classification System</td> <td style="border-bottom: 1px solid black;">Classification Symbols</td> </tr> <tr> <td style="height: 40px; vertical-align: bottom;">IPC5</td> <td style="height: 40px; vertical-align: bottom;">G 01 N</td> </tr> </table> <div style="text-align: center; border-top: 1px solid black; border-bottom: 1px solid black;">Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in Fields Searched⁸</div>			Classification System	Classification Symbols	IPC5	G 01 N
Classification System	Classification Symbols					
IPC5	G 01 N					
III. DOCUMENTS CONSIDERED TO BE RELEVANT⁹						
Category *	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³				
A	EP, A2, 0314525 (MAK, ERWIN A.) 3 May 1989, see column 5, line 7 - line 30; figures 1-3 --	1-21				
A	EP, A2, 0356250 (EASTMAN KODAK COMPANY) 28 February 1990, see figures 1-6 --	1-21				
A	GB, A, 2199407 (GENETIC SYSTEMS CORPORATION) 6 July 1988, see figures 1-5,11 --	1-21				
A	GB, A, 2239093 (KABUSHIKI KAISHA NITTEC) 19 June 1991, see page 12, line 11 - line 17; page 13, line 13 - page 15, line 14; page 17, line 6 - page 18, line 43; figures 1,2,6 --	1-21				
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>* Special categories of cited documents:¹⁰</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"&" document member of the same patent family</p> </div> </div>						
IV. CERTIFICATION						
Date of the Actual Completion of the International Search 4th November 1992		Date of Mailing of this International Search Report <div style="text-align: center; font-size: 1.2em;">13 NOV 1992</div>				
International Searching Authority <div style="text-align: center; font-weight: bold;">EUROPEAN PATENT OFFICE</div>		Signature of Authorized Officer <div style="text-align: center; font-weight: bold;">Nils Engnell</div>				

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No
A	FR, A1, 2591344 ("SO.F.I.S." SOCIÉTÉ FRANCAISE D'INSTRUMENTATION SCIENTIFIQUE) 12 June 1987, see page 6, line 14 - page 7, line 9; figures 4,5 --	1-21
A	Patent Abstracts of Japan, Vol 13, No 87, P835, abstract of JP 63-271164, publ 1988-11-09 (SHIMADZU CORP) -- -----	1-21

**ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO. PCT/US 92/06058**

SA 62971

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 30/09/92. The European Patent office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP-A2- 0314525	03/05/89	JP-A- 1153999	16/06/89
EP-A2- 0356250	28/02/90	US-A- 5008082	16/04/91
GB-A- 2199407	06/07/88	AU-B- 613881	15/08/91
		AU-D- 8040787	05/05/88
		BE-A- 1001048	20/06/89
		CH-A- 675635	15/10/90
		DE-A- 3736632	19/05/88
		FR-A- 2606156	06/05/88
		JP-A- 63259468	26/10/88
		LU-A- 87032	08/05/89
		NL-A- 8702591	16/05/88
		SE-A- 8704230	01/05/88
		US-A- 5096670	17/03/92
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		JP-A- 3208278	11/09/91
		JP-A- 3210883	13/09/91
		JP-A- 3212875	18/09/91
		JP-A- 3231205	15/10/91
		JP-A- 3175361	30/07/91
FR-A1- 2591344	12/06/87	NONE	

For more details about this annex : see Official Journal of the European patent Office, No. 12/82